



## Review Article

# Avicenna's Canon of Medicine and Tuberculosis: A Review on Herbal Medicine in Animal Model Research

Zakiyeh Sakhavat Nia<sup>1,2</sup> , Mehdi Sobhani<sup>3</sup> , and Zahra Sobhani<sup>1\*</sup>

<sup>1</sup> Department of Traditional Pharmacy, Faculty of Pharmacy, Mashhad University of Medical Sciences, Mashhad, Iran

<sup>2</sup> Student Research Committee, Mashhad University of Medical Sciences, Mashhad, Iran

<sup>3</sup> Endoscopic and Minimally Invasive Surgery Research Center, Mashhad University of Medical Sciences, Mashhad, Iran

\* **Corresponding author:** Zahra Sobhani, Department of Traditional Pharmacy, Faculty of Pharmacy, Mashhad University of Medical Sciences, Mashhad, Iran, Email: Sobhaniz@mums.ac.ir

### ARTICLE INFO

#### Article History:

Received: 20/08/2025

Revised: 29/09/2025

Accepted: 17/10/2025

Published: 30/10/2025



#### Keywords:

Anti-inflammatory

Antioxidant

Avicenna

Bioactive compound

*Mycobacterium tuberculosis*

### ABSTRACT

Tuberculosis (TB) remains a major global health challenge, highlighting the need for new and complementary therapeutic methods and strategies. The present study aimed to provide a comprehensive review of medicinal plants recommended by the renowned Persian physician Avicenna (Ibn Sina) for TB treatment, focusing on their phytochemical compounds and mechanisms of action. The present study combined a historical analysis of Avicenna's Canon of Medicine to identify medicinal plants used for tuberculosis with a systematic literature review (2000-2024) to evaluate their modern pharmacological evidence. The study targeted antimycobacterial, immunomodulatory, and symptom-relief activities using databases including PubMed, Scopus, and Science Direct. The current findings indicated that several plants, including *Artemisia absinthium* L., *Artemisia vulgaris* L., *Glycyrrhiza glabra* L., *Hyssopus officinalis* L., *Myrtus communis* L., *Thymus vulgaris* L., *Rosa damascena* Mill., *Adiantum capillus-veneris* L., *Achillea millefolium* L., *Foeniculum vulgare* Mill., *Polygonum aviculare* L., *Phoenix dactylifera* L., and *Teucrium polium* L., have multifaceted approaches against TB through potent anti-inflammatory, antioxidant, immunomodulatory, and direct antimycobacterial effects. Bioactive compounds included in these plants, such as phenolic acids, flavonoids, and terpenoids, are identified as key contributors that reduce oxidative stress, modulate immune responses, inhibit inflammatory mediators such as Interleukin-6, Interleukin-1 $\beta$ , and Tumor Necrosis Factor-alpha, and directly suppress *Mycobacterium tuberculosis* growth. Furthermore, these compounds help mitigate pulmonary damage and enhance host immune defenses. By integrating Avicenna's traditional knowledge with contemporary pharmacological evidence, the potential of these plants as complementary therapeutic agents for TB was noted.

## 1. Introduction

Tuberculosis (TB), caused by *Mycobacterium tuberculosis* (*M. tuberculosis*), is a significant global health threat, which mainly affects the respiratory system but can also impact other organs such as the kidneys and spine<sup>1</sup>. Transmitted via airborne particles, it is a disease of antiquity that persists as a leading infectious cause of mortality worldwide, with the World Health Organization (WHO) reporting an estimated 10 million new cases and 1.6 million deaths in 2021<sup>2</sup>. The emergence of multidrug-resistant tuberculosis (MDR-TB) and extensively drug-resistant tuberculosis (XDR-TB) strains presents a major challenge to treatment, underscoring the urgent need for novel

complementary and alternative therapeutic strategies<sup>3</sup>.

In response to the challenges of TB treatment, particularly drug-resistant strains, researchers are increasingly re-evaluating traditional herbal remedies. Before antibiotics, many cultures used medicinal plants to manage TB symptoms and slow disease progression, as documented in various traditional medical systems. Given the rise of drug resistance and the limitations of current treatments, the scientific study of these natural compounds may yield novel therapeutic strategies. Modern pharmacology now places significant emphasis on investigating bioactive compounds from plants with a history of medicinal use<sup>4</sup>. Validating these traditional

Cite this paper as: Sakhavat Nia Z, Sobhani M, and Sobhani Z. Avicenna's Canon of Medicine and Tuberculosis: A Review on Herbal Medicine in Animal Model Research. Journal of Lab Animal Research. 2025; 4(5): 43-56. DOI: 10.58803/jlar.v4i5.83



The Author(s). Published by Rovedar. This is an open-access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

approaches through rigorous science not only utilizes this ancient medical legacy but may also discover complementary or alternative therapies for conditions including drug-resistant TB<sup>5</sup>.

Avicenna (Ibn Sina), the preeminent 10th-11th century Persian physician, synthesized Greek and Persian knowledge to create a comprehensive medical system based on observation and scientific analysis<sup>6</sup>. His seminal work, *The Canon of Medicine*, became a foundational text for centuries, detailing disease classification, diagnostics, and the extensive use of medicinal plants<sup>7</sup>. By emphasizing empirical evidence and a holistic approach, Avicenna established scientific principles that continue to hold relevance in modern medicine<sup>6,7</sup>.

Avicenna's significant contributions to herbal medicine remain relevant today. While his principles were based on humoral theory, his detailed use of botanical treatments provides a valuable foundation for modern phytotherapy. In *The Canon of Medicine*, he identified specific plants for managing tuberculosis. Reviewing his recommendations can illuminate the potential of plant-based treatments, especially against antibiotic-resistant strains, highlighting the need for rigorous scientific research to validate their efficacy and safety for modern clinical use<sup>8</sup>. The study examines Avicenna's plant-based treatments for tuberculosis in *The Canon of Medicine* and compares them with modern scientific evidence to assess their potential value as complementary therapies.

## 2. Materials and Methods

### 2.1. Search strategy

Medicinal plants used for the treatment of TB were extracted from volume 3 of Avicenna's *Canon of Medicine* (*Al-Qanun fi al-Tibb*), specifically the tenth Treatise (Fann 10), fourth article (Maqala 4). A manual review was conducted using Arabic terms such as السِّل (al-Sill), داء السِّل (TB disease), and علاج السِّل بالأعشاب (herbal treatment of TB). Identified plant names were transliterated and subsequently matched with validated modern botanical nomenclature using reference floras and an ethnopharmacological database.

### 2.2. Systematic search in scientific databases

A systematic literature search was performed across the following electronic databases: PubMed, Scopus, Science Direct, Cochrane Library, and Google Scholar as a supplementary tool to identify grey literature and articles not indexed in the core databases, covering the period from January 2000 to March 2024. The literature search was conducted by using a combination of plant names, medical subject terms, and keywords associated with therapeutic outcomes. Boolean operators (AND/OR) were used to refine the queries. The following keyword clusters were applied. Antitubercular activity, Antitubercular or Antimycobacterial or *Mycobacterium tuberculosis* inhibition, Symptom relief, Antitussive or Antipyretic or Analgesic or Antifatigue, pulmonary protective effects, Bronchodilator or Anti-asthmatic or Pulmonary fibrosis or Lung injury. For adjunctive and immunological effects, Immunostimulant, Anti-inflammatory, Antioxidant, or Antibacterial were searched. Examples of search query syntax for a single plant species were *Glycyrrhiza glabra* or Licorice and

Antitubercular or Antioxidant or Immunomodulatory or Bronchodilator or Anti-inflammatory.

### 2.3. Inclusion criteria

Articles published in English from 2000 to 2024. *In vitro*, *in vivo*, or clinical studies evaluating the pharmacological activity of the selected plant species in the context of TB or its symptoms, studies reporting mechanisms of action, pharmacodynamics, or synergistic effects.

### 2.4. Exclusion criteria

Studies unrelated to tuberculosis or lung diseases, reviews without original data reports, and lacking botanical authentication.

## 3. Etiology and epidemiology of tuberculosis

Tuberculosis remains a highly prevalent and lethal global health challenge, disproportionately affecting low- and middle-income countries where factors such as poverty and inadequate sanitation facilitate its spread. However, it persists as a public health issue even in high-income nations. The COVID-19 pandemic exacerbated the TB crisis by disrupting essential diagnostic and treatment services globally. According to the 2024 WHO report, an estimated 10.8 million people were diagnosed with TB in 2023, a slight increase from the previous year. Although TB-related deaths dropped to 1.25 million in 2023, the disease has likely regained its position as the world's leading cause of death from a single infectious agent, surpassing COVID-19<sup>9,10</sup>.

Tuberculosis progresses through two distinct clinical stages, including latent TB infection (LTBI) and active TB disease<sup>11</sup>. In LTBI, which constitutes about 90% of all TB infections, individuals harbor dormant *M. tuberculosis* bacteria, are asymptomatic, and non-infectious, though the infection can reactivate later, particularly if the immune system becomes compromised<sup>11,12</sup>. Active TB disease occurs when the bacteria reactivate and multiply, causing symptoms such as a chronic cough, weight loss, fever, and hemoptysis. Unlike LTBI, individuals with active pulmonary TB are contagious and can transmit the bacterium to others<sup>11</sup>.

Pulmonary tuberculosis follows a defined pathogenic sequence transmitted through aerosol inhalation. The process initiates when immune cells phagocytose the bacteria, followed by intracellular bacterial replication. The infection may then enter a latent, asymptomatic phase, where it can persist indefinitely. Ultimately, factors such as immunosuppression can trigger reactivation to active disease, characterized by symptomatic illness and transmission potential<sup>13</sup>.

### 3.1. Transmission and action mechanisms

The infectiousness of a TB patient depends on several factors. Those with smear-positive pulmonary TB are highly contagious, and the risk increases with the degree of smear positivity<sup>14</sup>. A study in Peru confirmed that smear-positive individuals pose a significantly higher transmission risk to household contacts than smear-negative cases, regardless of the contact's age<sup>15,16</sup>. *Mycobacterium tuberculosis* transmits through airborne droplets expelled by infected individuals via coughing or sneezing. When inhaled, the bacteria travel to and establish infection in the alveoli of the lungs<sup>17</sup>. The host-

pathogen interaction in tuberculosis is complex. The initial innate immune response recruits macrophages to contain *M. tuberculosis*, but this often fails, leading to granuloma formation. This structure, while intended to wall off the infection, paradoxically shelters the bacteria, enabling long-term persistence<sup>17</sup>. This homeostatic balance is vulnerable; immunosuppression, such as HIV, significantly increases the risk of active TB, highlighting the direct correlation between immune status and disease progression<sup>18,19</sup>. The process involves sophisticated co-evolution. After initial infection and innate response in the lungs, *M. tuberculosis* migrates to lymph nodes to prime T-cells<sup>20,21</sup>. These activated immune cells then travel back to the lungs, reinforcing granuloma formation where *M. tuberculosis* can establish a latent, protected state. Dendritic cells and macrophages serve as the primary defense against TB, working in concert to control bacterial replication<sup>22,23</sup>. Dendritic cells and macrophages form granulomas, organized structures of macrophages, giant cells, and lymphocytes, which paradoxically create a niche for *M. tuberculosis* survival. The stability of this structure is regulated by cytokines, including the pro-inflammatory ones, such as interleukin 1- $\beta$  (IL-1 $\beta$ ), interferon gamma (IFN- $\gamma$ ),

and tumor necrosis factor-alpha (TNF- $\alpha$ ), which enhance immune function, while anti-inflammatory ones, such as IL-10, maintain homeostasis<sup>21,22</sup>. In about 10% of cases, *M. tuberculosis* evades this containment, causing granulomas to break down. This marks the progression from latent infection to active, symptomatic, and transmissible disease<sup>24</sup>.

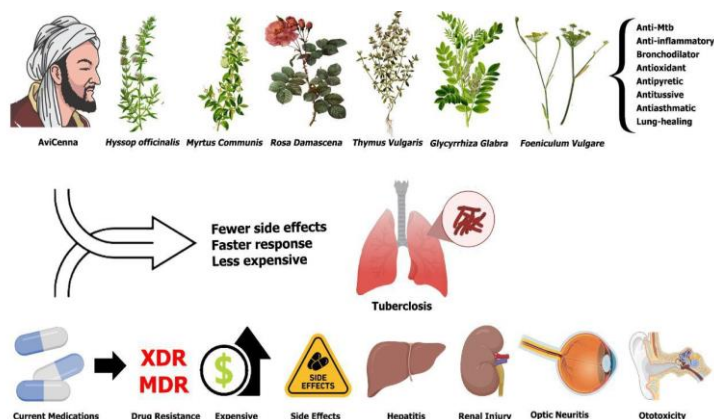
#### 4. Avicenna's perspective on tuberculosis

The renowned Persian physician Avicenna employed a holistic approach to treating TB, combining botanical remedies, nutritional adjustments, and other therapies to strengthen the immune system and alleviate symptoms<sup>25,26</sup>. Many of the medicinal plants he recommended possess direct antibacterial activity against *M. tuberculosis* while also providing relief from symptoms like fever and cough. These natural supplements are recognized for improving patient conditions and reducing disease severity. This section investigates the therapeutic impact of Avicenna's recommended herbal remedies on TB<sup>25,26</sup>. The summary of medicinal plants recommended by Avicenna for tuberculosis is mentioned in Table 1 and Figure 1.

**Table 1.** Summary of medicinal plants recommended by Avicenna for treating tuberculosis

Medicinal plants	Key mechanisms of action	Reference numbers
<i>Artemisia absinthium</i> L.	Anti-inflammatory, antioxidant, antipyretic, direct bacteriostatic/bactericidal effects against <i>M. tuberculosis</i>	(32-38)
<i>Artemisia vulgaris</i> L.	Anti-inflammatory, antioxidant, bronchodilator, anti-asthmatic, antibacterial against <i>M. tuberculosis</i>	(39-47)
<i>Glycyrrhiza glabra</i> L.	Anti-inflammatory, antioxidant, immunomodulatory, antitussive, anti-asthmatic, hepatoprotective, anti-fatigue, protects against lung injury	(48-65)
<i>Hyssopus officinalis</i> L.	Anti-inflammatory, antioxidant, anti-asthmatic, modulates cytokines	(67-73)
<i>Myrtus communis</i> L.	Anti-inflammatory, antioxidant, and direct antimicrobial activity against <i>M. tuberculosis</i>	(71, 74-81)
<i>Thymus vulgaris</i> L.	Anti-inflammatory, antioxidant, bronchodilator, anti-asthmatic, and direct antimycobacterial activity	(82-95)
<i>Rosa damascena</i> Mill.	Anti-inflammatory, antioxidant, bronchodilator	(96-103)
<i>Adiantum capillus-veneris</i> L.	Anti-inflammatory, antioxidant, antipyretic, anti-apoptotic (lung protection), hepatoprotective	(104-115)
<i>Achillea millefolium</i> L.	Anti-inflammatory, antioxidant, immunomodulatory, bronchodilator (Tracheal relaxant)	(116-126)
<i>Foeniculum vulgare</i> Mill.	Anti-inflammatory, antioxidant, bronchodilator, direct anti-TB activity	(127-136)
<i>Polygonum aviculare</i> L.	Anti-inflammatory, antioxidant, anti-fatigue (modulates neuroinflammation)	(137-145)
<i>Phoenix dactylifera</i> L.	Anti-inflammatory, antioxidant, immunomodulatory, protects against lung injury and fibrosis	(146-157)
<i>Teucrium polium</i> L.	Anti-inflammatory, antioxidant, immunomodulatory	(158-167)

*M. tuberculosis*: *Mycobacterium tuberculosis*



**Figure 1.** An overview of medicinal plants with anti-inflammatory, antioxidant, bronchodilatory, and anti-tuberculosis properties, drawing on traditional medicine and Avicenna's teachings to highlight their potential for fewer side effects, faster therapeutic response, and lower cost compared to conventional tuberculosis treatments.

##### 4.1. *Artemisia absinthium* L.

*Artemisia* L. is a plant that grows in northern arid areas.

*Artemisia* L. has 1000 genera and more than 20000 species found in different regions in the world, including Asia, Europe, and North America<sup>29</sup>. *Artemisia* has several active

ingredients and secondary metabolites that have a wide range of bioactive activities<sup>29</sup>. There are some fundamental classes of bioactive compounds found in *Artemisia*, including flavonoids, phenolic acids, coumarin, and terpenes<sup>30</sup>. *Artemisia* is mostly used for treating malaria, hepatitis, cancer, and different diseases caused by fungi, bacteria, or viruses<sup>29</sup>. These diseases are treatable owing to *Artemisia*'s diverse antitumor, anti-inflammatory, and antiulcer secondary metabolites<sup>29,30</sup>.

*Artemisia absinthium* L. (Asteraceae family) is a plant traditionally used to treat different ailments, including liver disorders, gastric pain, and wounds<sup>29</sup>. It exhibits broad pharmacological activities, such as antimicrobial, antioxidant, and hepatoprotective effects<sup>30</sup>. Notably, prolonged use of its essential oil is associated with toxicity and psychological dysfunction<sup>31</sup>. While the precise mechanisms of its antituberculosis activity are not fully understood, it is attributed to its diverse bioactive compounds.

#### 4.1.1 Anti-inflammatory effects

Studies have confirmed the anti-inflammatory properties of *A. absinthium*. Amrollahi et al.<sup>32</sup> demonstrated that its essential oil (4-8 mg/kg) significantly reduced carrageenan-induced paw edema in mice, with efficacy similar to aspirin<sup>32</sup>. Furthermore, *in vitro* tests exhibited that the plant has potent lipoxygenase inhibitory activity, comparable to standard anti-inflammatory medicines such as ibuprofen<sup>33</sup>.

#### 4.1.2. Antioxidant activities

*Artemisia absinthium* L. demonstrated significant antioxidant and liver-protecting properties. Craciunescu et al.<sup>34</sup> indicated that flavonoid-rich extracts have potent antioxidant activity in 2,2-Diphenyl-1-picrylhydrazyl (DPPH), Oxygen Radical Absorbance Capacity (ORAC), and Trolox Equivalent Antioxidant Capacity (TEAC) assays and protect cells from oxidative stress<sup>34</sup>. Supporting this, Amat et al.<sup>35</sup> found that its aqueous extract (50-200 mg/kg) in mice reduced liver enzymes and lipid peroxidation while restoring key antioxidant enzymes such as superoxide dismutase (SOD) and glutathione peroxidase (GPx). It also suppressed pro-inflammatory cytokines (TNF- $\alpha$ , IL-1) and prevented liver necrosis, confirming its hepatoprotective and antioxidant effects<sup>35</sup>.

#### 4.1.3. Antipyretic effects

*Artemisia absinthium* L. has demonstrated significant antipyretic (fever-reducing) properties. A study by Bhat et al.<sup>37</sup> induced fever in rats and treated them with a hydroalcoholic extract of the plant. The results indicated that the extract, both alone and combined with barley water, significantly reduced rectal temperature, confirming its anti-fever effects<sup>36,37</sup>.

#### 4.1.4. Antituberculosis effects

Given the hepatotoxicity of current TB medicines and the rise of MDR/XDR strains, exploring new agents such as medicinal plants is crucial. In one study, *A. absinthium* extracts showed dose-dependent anti-TB activity in animal models. All concentrations (20%-96%) had

bacteriostatic effects, but only the 96% extract was bactericidal, highlighting its potential as a therapeutic candidate that requires further investigation<sup>38</sup>.

#### 4.2. *Artemisia vulgaris* L.

*Artemisia vulgaris* L., a prominent medicinal plant within the *Artemisia* genus, is well recognized for its essential oils, which are abundant in compounds such as  $\alpha$ -thujone, camphor, and 1,8-cineole<sup>39,40</sup>. It possesses a wide range of pharmacological properties, including anti-inflammatory, antioxidant, and immunomodulatory effects, and has demonstrated significant antibacterial activity against *M. tuberculosis*<sup>41</sup>. These therapeutic effects are attributed to its diverse secondary metabolites, such as flavonoids, sesquiterpenes, and coumarins<sup>39,40,42</sup>.

##### 4.2.1. Anti-inflammatory effects

*Artemisia vulgaris* L. exhibits significant anti-inflammatory activity, partly attributed to its camphor content, which inhibits pro-inflammatory cytokines such as TNF- $\alpha$ , IL-2, and IL-4<sup>43</sup>. *In vivo* studies confirmed that methanolic extract (200-800 mg/mL) potently inhibited carrageenan-induced paw edema in rats by over 71%<sup>44</sup>. Another study found that ethanol extracts from plants in temperate regions (at 400 mg/kg) were as effective as standard anti-inflammatory medicines in a mouse model, with ethanol being a superior solvent to ethyl acetate<sup>43</sup>.

##### 4.2.2. Antioxidant activities

*Artemisia vulgaris* L. demonstrated potent antioxidant activity through both *in vitro* and *in vivo* studies. Temraz et al.<sup>45</sup> reported strong free radical scavenging in DPPH and Nitric Oxide assays, with treatment in rats increasing key antioxidant markers such as glutathione, ascorbic acid, and SOD<sup>45</sup>. Similarly, its aqueous extract effectively scavenged 2,2'-Azino-bis(3-ethylbenzothiazoline-6-sulfonic acid), superoxide, hydroxyl, and NO radicals, as shown by low IC<sub>50</sub> values<sup>46</sup>.

##### 4.2.3. Bronchodilator effects

*Artemisia vulgaris* L. acts as an effective bronchodilator. A study on guinea pig tracheal tissue exhibited that its extract potently prevented contractions induced by carbachol and potassium. The mechanism involves blocking muscarinic receptors and inhibiting calcium influx, leading to tracheal relaxation<sup>47</sup>.

#### 4.3. *Glycyrrhiza glabra* L.

*Glycyrrhiza glabra* (*G. glabra*, licorice), a species rich in bioactive compounds such as the triterpenoid saponin glycyrrhizin, flavonoids, and coumarins, possesses a broad spectrum of therapeutic properties, including anti-tuberculosis activity<sup>48,49</sup>.

##### 4.3.1. Anti-inflammatory effects

*Glycyrrhiza glabra* exhibits potent anti-inflammatory effects through its flavonoid constituents. Sun et al.<sup>50</sup> reported that liquiritigenin reduces inflammation by modulating MAPK and Notch1/NF- $\kappa$ B pathways. Another study identified

pinocembrin, glabranin, and licoflavanone as key anti-inflammatory compounds in *G. glabra*. In lipopolysaccharide (LPS)-stimulated macrophages, these compounds significantly decreased pro-inflammatory cytokines and COX-2/iNOS expression by inhibiting the NF- $\kappa$ B/MAPK pathway, with licoflavanone (IC<sub>50</sub> = 37.68  $\mu$ M) showing the strongest effect<sup>51</sup>.

#### 4.3.2. Anti-asthmatic effects

Glycyrrhizic acid, a primary active compound in *G. glabra*, demonstrated significant anti-asthmatic and immunomodulatory effects in mouse models. Studies indicated that *G. glabra* reduces key asthma indicators such as airway resistance, eosinophil count, and pro-inflammatory cytokines (IL-4, IL-5, IL-13), while boosting regulatory T cells and IFN- $\gamma$  levels<sup>52,53</sup>. These findings suggested that *G. glabra* may be effective against TB by mitigating lung inflammation and modulating the immune response, highlighting its potential as a therapeutic agent for respiratory diseases, such as TB.

#### 4.3.3. Antitussive effects

*Glycyrrhiza glabra* exhibits significant antitussive (cough-suppressing) and expectorant properties. Studies in mice and guinea pigs using different cough-inducing models (SO<sub>2</sub>, ammonia) have demonstrated that its extract and specific compounds, such as liquiritigenin, offer cough suppression that is comparable to or exceeds the efficacy of standard medicinal treatments codeine<sup>54-57</sup>. These findings confirmed its potential as an effective natural cough remedy.

#### 4.3.4. Antioxidant activities

*Glycyrrhiza glabra* flavonoid-rich root fractions possess potent antioxidant activity. A methanol/water extract demonstrated potent effects in multiple assays, particularly in inhibiting lipid peroxidation<sup>58</sup>. Furthermore, the root extract exhibits significant hepatoprotective properties. In a mouse model of CCl<sub>4</sub>-induced liver damage, pre-treatment with the extract (300-600 mg/kg) effectively reduced oxidative stress and protected against hepatotoxicity<sup>59</sup>.

#### 4.3.5. Fatigue relief effects

Fatigue is a common and debilitating symptom in TB, stemming from both the disease and its long-term treatments, which severely impacts patients' quality of life. Studies suggested *G. glabra* may alleviate fatigue. In mouse models, the compound glabridin enhanced exercise endurance, reduced fatigue markers such as blood lactic acid, and promoted glycogen storage<sup>60</sup>. Similarly, *G. glabra* extract effectively countered signs of induced chronic fatigue stress, indicating its potential as a therapeutic agent for fatigue management<sup>61</sup>.

#### 4.3.6. Treatment of lung injury

Lung damage, a hallmark of TB, ranges from acute inflammation and abscesses to chronic fibrosis, leading to permanent respiratory impairment. Glycyrrhizin, a key compound in *G. glabra*, demonstrated notable protective effects against such injury. In mouse models of LPS-induced

acute lung injury, glycyrrhizin treatment suppressed key inflammatory mediators (IL-6, TNF- $\alpha$ , IL-1 $\beta$ ) and inhibited the NF- $\kappa$ B/NLRP3 inflammasome pathway, thereby reducing lung inflammation and damage<sup>62,63</sup>. These findings suggested its potential to mitigate TB-associated lung injury.

#### 4.3.7. Antituberculosis effects

Several studies demonstrated the direct therapeutic potential of *G. glabra* against TB. An inhalable liposomal powder of licorice extract was effectively deposited in mouse lungs, significantly reducing bacterial counts in the lungs and spleen over 56 days<sup>64</sup>. Furthermore, a clinical trial on TB patients found that adding licorice as an adjunct therapy for eight weeks improved sputum conversion rates by 10%, resolved fever more effectively, provided more effective cough relief, and resulted in fewer gastrointestinal side effects compared to the placebo group<sup>65</sup>.

### 4.4. *Hyssopus officinalis* L.

Hyssop, a perennial herb belonging to the *Lamiaceae* family, has been historically valued for its diverse applications in both medicine and culinary practices. Extracts and essential oils derived from hyssop are used for anti-inflammatory, antioxidant, antibacterial, antiviral, expectorant, antiseptic, carminative, tonic, diuretic, and cardiovascular effects<sup>66</sup>.

#### 4.4.1. Anti-inflammatory effects

Hyssop exhibits significant anti-inflammatory activity. An ethanolic extract reduced inflammation in a rat paw edema model by lowering phagocyte levels and nitrite/nitrate<sup>67</sup>. Another study found that a methanol extract (200 mg/kg) inhibited paw edema as effectively as indomethacin. *In vitro*, the extract potently inhibited the COX-2 enzyme, comparable to celecoxib. Molecular modeling identified chlorogenic acid and rosmarinic acid as key compounds with strong binding affinity to COX-1 and COX-2, surpassing ibuprofen<sup>68</sup>.

#### 4.4.2. Anti-asthmatic effects

Hyssop demonstrated remarkable anti-asthmatic potential. In an ovalbumin-induced asthma model in mice, hyssop treatment effectively normalized key asthma indicators. It reduced elevated eosinophil and immunoglobulin E (IgE) levels, increased IgG, and decreased excessive airway mucus secretion<sup>69</sup>. Furthermore, the treatment reduced levels of matrix metalloproteinase-9 and its inhibitor, mitigating collagen deposition and mucus hypersecretion, which supports its role in preventing airway remodeling in asthma<sup>70</sup>.

#### 4.4.3. Antioxidant activities

Hyssop exhibits notable antioxidant activity across its flowers, leaves, and stems. Studies using DPPH radical scavenging and lipid peroxidation assays confirm this effect, with one methanolic extract showing 40% inhibition of lipid peroxidation<sup>71,72</sup>. This antioxidant capacity is directly correlated with the plant's phenolic content<sup>73</sup>.

#### 4.5. *Myrtus communis* L.

*Myrtus communis* L. (myrtle), an aromatic evergreen shrub, has been used in traditional medicine to treat coughs, diarrhea, and for its antimicrobial properties<sup>74,75</sup>. Its specific mechanisms of action against TB are discussed next.

##### 4.5.1. Anti-inflammatory effects

*Myrtus communis* L. possesses potent anti-inflammatory properties. In a rat model of colitis, both its hydroalcoholic extract and essential oil effectively reduced inflammation, as shown by improved ulcer index, colitis index, and myeloperoxidase activity<sup>77</sup>. The activity is attributed to specific compounds, such as myrtucommulone, which directly inhibits key enzymes (COX-1, 5-lipoxygenase) in the inflammatory pathway<sup>78</sup>. In mice, myrtucommulone reduced inflammation by lowering leukocyte infiltration, myeloperoxidase activity, and pro-inflammatory cytokines (TNF- $\alpha$ , IL-1 $\beta$ )<sup>79</sup>.

##### 4.5.2. Antioxidant activities

*Myrtus communis* L. exhibits strong antioxidant activity, primarily attributed to its high phenolic content. Methanolic extracts from its leaves, stems, and flowers indicated superior antioxidant effects compared to essential oils, with leaves being the most potent part<sup>80,81</sup>. The main antioxidants identified are gallotannins in leaves and flowers, and catechin in stems<sup>80</sup>. Methanol is the most effective solvent for extracting these antioxidant compounds<sup>81</sup>.

##### 4.5.3. Antituberculosis effects

The essential oil of myrtle demonstrated significant and selective antimicrobial activity. It is effective against *M. tuberculosis* strains, such as H37Rv, H37Ra, and multi-drug resistant clinical isolates, with a low MIC of 0.17% (v/v). In contrast, myrtle was far less effective against *M. paratuberculosis* (MIC of 2%)<sup>81</sup>. The anti-bacterial mechanism against gram-positive bacteria is reported to involve cell wall destruction.

#### 4.6. *Thymus vulgaris* L.

*Thymus vulgaris* L. (*T. vulgaris*, thyme), a fragrant shrub from the *Lamiaceae* family, is traditionally used to treat respiratory ailments such as bronchitis, coughs, and sore throats<sup>82</sup>. Its essential oil possesses broad antimicrobial, antiviral, and antioxidant properties<sup>83</sup>. These effects are primarily due to phenolic monoterpenes, including thymol and carvacrol, which are also potent antifungals. Other constituents may act synergistically to enhance the oil's overall efficacy<sup>84-86</sup>.

##### 4.6.1. Anti-inflammatory effects

Thyme essential oil and its constituents, thymol and carvacrol, exhibit anti-inflammatory activity, but their effects vary. In a pleurisy model, both thymol and carvacrol inhibited leukocyte migration. However, in a topical ear edema test, only carvacrol was effective, while thymol caused irritation, possibly by triggering prostanoid and histamine release<sup>87</sup>.

Additionally, both thyme and thymol can modulate inflammation by suppressing the gene expression and production of key pro-inflammatory cytokines, including IL-6, IL-1 $\beta$ , and TNF- $\alpha$ <sup>88</sup>.

##### 4.6.2. Anti-asthmatic effects

Thyme oil demonstrates significant anti-asthmatic and antioxidant effects. In an ovalbumin-induced asthma model in rabbits, thyme oil treatment normalized lung tissue, reduced signs (wheezing, sneezing), and lowered key Th2 cytokines (IL-4, IL-5, IL-13), IgE, and reactive oxygen species (ROS)<sup>89</sup>. The antioxidant properties of thyme and its constituent thymol have been further confirmed by their ability to reduce free radicals in mice<sup>90</sup>.

##### 4.6.3. Antioxidant activities

As reported in the previous section, thyme exhibits therapeutic activity against bronchial asthma through various mechanisms, particularly through its antioxidant properties<sup>89,90</sup>. Additionally, *T. vulgaris* silver nanoparticles demonstrated significant DPPH radical scavenging activity in a study by Aldosary et al.<sup>91</sup>.

##### 4.6.4. Antituberculosis effect

*Thymus vulgaris* L. exhibits potent and direct activity against *M. tuberculosis*. Its extracts and essential oil demonstrated notable growth inhibition, with studies reporting low MIC values (0.5 mg/ml for acetone extract and as low as 0.5-40  $\mu$ g/ml for essential oil)<sup>92,93</sup>. Notably, the essential oil indicated bactericidal effects at a concentration of 1  $\mu$ g/ml<sup>94</sup>, and its efficacy has been found comparable to, or even surpassing, that of standard anti-TB medicines such as isoniazid and streptomycin in some assays<sup>95</sup>.

#### 4.7. *Rosa damascena* L.

*Rosa damascena* L. (*R. damascena*) is one of the most important aromatic species of the *Rosaceae* family. Its essential oil is highly valued in perfumery and cosmetics. Numerous therapeutic activities of *R. damascena* have been reported, including its analgesic properties, anti-muscle cramp effects, anti-inflammatory actions, anti-cough properties, antioxidant activity, and anti-cancer effects<sup>96</sup>.

##### 4.7.1. Anti-inflammatory effects

*Rosa damascena* essential oil exhibited remarkable anti-inflammatory and antioxidant properties. In a rat model of sepsis-induced inflammation, pre-treatment with the oil (50-100 mg/kg) prevented hepatotoxicity by reducing oxidative stress, lipid peroxidation, and modulating key inflammatory mediators, such as COX-2 and PGE2<sup>97,98</sup>. These findings suggested its potential as an alternative to conventional anti-inflammatory medicines.

##### 4.7.2. Antioxidant activities

*Rosa damascena* demonstrated potent antioxidant activity, primarily attributed to its high content of phenolic compounds, flavonoids, anthocyanins, quercetin, and kaempferol<sup>99</sup>. This is confirmed by strong performance in standard assays, including DPPH, Ferric Reducing



Antioxidant Power (FRAP), ABTS, with significant activity observed across different plant accessions<sup>100</sup>. The petal extract is notably more potent than the receptacle extract, showing effectiveness at much lower concentrations<sup>101</sup>.

#### 4.7.3. Bronchodilator activity

*Rosa damascena* L. acts as an effective bronchodilator. Its extracts and essential oil induce dose-dependent relaxation of tracheal smooth muscle in animal models. The mechanism is multi-faceted, involving potential stimulation of  $\beta$ -adrenoceptors, blockade of histamine (H1) receptors<sup>102</sup>, and the inhibition of multiple potassium channels (voltage-gated, ATP-sensitive, and calcium-activated)<sup>103</sup>.

#### 4.8. *Adiantum capillus-veneris* L.

*Adiantum capillus-veneris* L., an evergreen fern, is traditionally used to treat respiratory ailments. Its essential oil, containing compounds such as carvacrol and thymol, exhibits documented antibacterial and antifungal properties<sup>104-106</sup>.

##### 4.8.1. Anti-inflammatory effects

*Adiantum capillus-veneris* L. demonstrated significant anti-inflammatory and hepatoprotective effects. In rat models of colitis, both its aqueous and hydroalcoholic extracts reduced inflammation in a dose-dependent manner, an effect attributed to their phenolic and flavonoid content<sup>107</sup>. Similarly, a methanolic extract notably reduced carrageenan-induced paw edema in mice<sup>108</sup>. Furthermore, the extract exhibited hepatoprotective properties in rats exposed to a toxic fungicide, reducing liver damage by lowering oxidative stress and pro-inflammatory cytokines (TNF- $\alpha$ , IL-6)<sup>109</sup>.

##### 4.8.2. Antioxidant activities

*Adiantum capillus-veneris* L. is a valuable source of antioxidants. Its activity is attributed to flavonoids, such as rutin and isoquercetin, which boost endogenous antioxidants (glutathione, catalase, SOD) and scavenge free radicals<sup>107,109</sup>. The essential oil, rich in carvone, carvacrol, and thymol, also exhibited strong DPPH radical scavenging ability<sup>110</sup>. Studies confirmed that methanol and ethyl acetate extracts have more potent antioxidant and enzyme-inhibitory effects than aqueous extracts<sup>111,112</sup>, collectively underscoring the plant's significant antioxidant potential.

##### 4.8.3. Antipyretic effects

*Adiantum capillus-veneris* exhibits significant antipyretic (fever-reducing) activity. In a mouse model, a methanolic extract (600 mg/kg) reduced yeast-induced fever more effectively than a standard dose of paracetamol, lowering rectal temperature from 38.63°C to 37.09°C within three hours<sup>108</sup>. This effect is likely due to a variety of phytochemicals, including flavonoids, terpenoids, and alkaloids present in the plant<sup>108</sup>.

##### 4.8.4. Treatment of lung injury

*Adiantum capillus-veneris* extract demonstrated protective effects against exercise and hypoxia-induced lung cell apoptosis. In trained rats under hypoxic conditions, the

extract (500 mg/kg) modulated key apoptotic regulators (P53, TNF- $\alpha$ , Bax/Bcl-2 ratio) and increased respiratory surface area<sup>113,114</sup>. Additionally, in interval-trained rats, an ethanol extract (200 mg/kg) acted as a pulmonary oxidative stress modulator by beneficially regulating metallothionein levels<sup>115</sup>.

#### 4.9. *Achillea millefolium* L.

*Achillea millefolium* L. (yarrow), a medicinal plant from the *Asteraceae* family, has been used for centuries to treat a wide range of ailments. These include liver disorders such as hepatitis, jaundice, respiratory issues including cough, pneumonia, fever, and inflammatory conditions, including rheumatoid arthritis, highlighting its broad pharmacological potential<sup>116, 117</sup>.

##### 4.9.1. Anti-inflammatory effects

*Achillea millefolium* L. exhibited potent anti-inflammatory properties through multiple mechanisms. *In vitro*, its extract suppresses key pro-inflammatory mediators (NO, iNOS, COX-2, IL-6, IL-8) in immune and skin cells, likely by inhibiting the NF- $\kappa$ B and p38 MAPK pathways<sup>118,119</sup>. *In vivo*, the extract effectively alleviated dermatitis signs in mice<sup>118</sup>. Its protective effect was further confirmed against toxins from *Clostridium difficile*, highlighting its broad anti-inflammatory and cell-protective potential<sup>120</sup>.

##### 4.9.2. Antioxidant activities

*Achillea millefolium* L. demonstrated significant antioxidant activity. Both its hydrodistilled essential oil (IC<sub>50</sub> = 1.83 mg/mL in DPPH assay) and hydroalcoholic extracts indicated potent free radical scavenging ability across multiple assays (DPPH, ABTS, CUPRAC, FRAP)<sup>121,122</sup>. Decoction extracts exhibit the highest activity, and the effects are largely attributed to bioactive compounds such as thymol and carvacrol<sup>122,123</sup>.

##### 4.9.3. Bronchodilator activity

The hexane extract from the aerial parts of *A. millefolium* L. demonstrated significant tracheal smooth muscle relaxant effects in rats (EC<sub>50</sub> = 412.0  $\mu$ g/mL). The mechanism involves muscarinic receptor antagonism, activation of the NO/cGMP pathway, and inhibition of calcium influx into smooth muscle cells. The compounds leucodin and achillin are identified as the key active constituents responsible for this bronchodilatory effect<sup>124</sup>.

##### 4.9.4. Immunomodulatory Effect

*Achillea millefolium* L. exhibited remarkable immunomodulatory properties. A methanolic extract increased lymphocyte and monocyte counts in mice, indicating an immunostimulatory effect attributed to its alkaloids, tannins, and flavonoids<sup>125</sup>. Conversely, the polysaccharide fraction of an aqueous extract demonstrated a more complex immunoregulatory role in human monocytes. It promoted the secretion of both pro- and anti-inflammatory cytokines (e.g., TNF- $\alpha$ , IL-10) while simultaneously inhibiting key signaling pathways, such as NF- $\kappa$ B, ERK1/2, and Akt<sup>126</sup>.

#### 4.10. *Foeniculum vulgare* Mill

*Foeniculum vulgare* Mill. (fennel), an *Apiaceae* family plant used as a spice is traditionally employed to relieve respiratory symptoms. It possesses broad pharmacological activities, including antibacterial, antioxidant, and hepatoprotective effects<sup>127,128</sup>.

##### 4.10.1. Anti-inflammatory effects

*Foeniculum vulgare* Mill exhibits significant anti-inflammatory activity. Its coumarin compounds, particularly imperatorin, inhibit pro-inflammatory cytokines (IL-6, TNF- $\alpha$ , IL-1 $\beta$ ) and COX-2 expression in both cellular and mouse models<sup>129</sup>. Similarly, a fennel extract suppressed IL-6 and TNF- $\alpha$  production in LPS-induced mice. The mechanism also involved modulation of NO levels, LDH activity, immune cell infiltration, and phosphorylation of the ERK signaling pathway<sup>130</sup>.

##### 4.10.2. Antioxidant activities

*Foeniculum vulgare* Mill. is a well-established source of antioxidants. Its coumarin compounds exhibit free radical scavenging activity in DPPH and ABTS assays<sup>129</sup>. Comparative studies on fennel seeds confirm significant antioxidant capacity across various assays (beta-carotene/linoleic acid, reducing power, DPPH), with some variation in potency observed between different geographical sources<sup>131,132</sup>.

##### 4.10.3. Bronchodilator effects

*Foeniculum vulgare* Mill. demonstrated bronchodilatory properties. Both its ethanolic extract and essential oil induce relaxation of guinea pig tracheal smooth muscle<sup>133</sup>. The compound fenchone, present in fennel, acts as a potent bronchodilator. Its mechanism primarily involves the activation of potassium (K<sup>+</sup>) channels, with additional contributions from phosphodiesterase inhibition and calcium (Ca<sup>2+</sup>) channel blockade<sup>134</sup>.

##### 4.10.4. Antituberculosis effect

*Foeniculum vulgare* Mill. exhibited direct antimicrobial and anti-tuberculosis activity. Different leaf extracts (petroleum ether, chloroform, methanol) showed efficacy against *M. tuberculosis*, with the chloroform extract being active in a TB-specific assay<sup>135</sup>. Furthermore, a hexane extract from *Foeniculum vulgare vardulce* was effective against MDR *M. tuberculosis* strains. Bioassay-guided fractionation identified 2,4-undecadienal as the most active compound, with a promising MIC of 25–50  $\mu\text{g/mL}$ <sup>136</sup>.

#### 4.11. *Polygonum aviculare* L.

*Polygonum aviculare* L. (knotgrass), a climbing plant from the *Polygonaceae* family, is traditionally used for its astringent (against diarrhea), diuretic, and healing properties<sup>137,138</sup>. Its activity is attributed to compounds, such as avicularin, myricitrin, and different diterpenes<sup>137,138</sup>.

##### 4.11.1. Anti-inflammatory effects

*Polygonum aviculare* L. demonstrated significant anti-inflammatory activity. Its isolated flavonol glucuronides

potently inhibit ROS production and elastase release in human neutrophils at low concentrations (0.5-10  $\mu\text{M}$ )<sup>139</sup>. Furthermore, the extract reduces DNA damage ( $\gamma\text{H2AX}$  formation) and modulates transcription factor activity, confirming its role as a potent inflammatory regulator<sup>140</sup>.

##### 4.11.2. Antioxidant activities

*Polygonum aviculare* L. is rich in phenolic antioxidants. An optimized deep eutectic solvent extraction method yielded extracts with strong DPPH and FRAP activity<sup>141,142</sup>. Comparative analysis shows that the methanolic extract of the stems possesses the highest antioxidant capacity, followed by the roots and leaves<sup>143</sup>.

##### 4.11.3. Fatigue relief effects

*Polygonum aviculare* L. extract demonstrated significant anti-fatigue effects. In a mouse model of stress-induced fatigue, treatment with the extract (100 mg/kg) reduced lethargy and modulated key physiological markers. Knotgrass lowered stress hormones, including corticosterone, epinephrine, neurotransmitters (serotonin), and pro-inflammatory cytokines (TNF- $\alpha$ , IL-1 $\beta$ , IL-6) in the brain, indicating that its anti-fatigue action works through the modulation of neuroinflammation<sup>144,145</sup>.

#### 4.12. *Phoenix dactylifera* L.

*Phoenix dactylifera* L. (date palm) is cultivated for its nutritious fruit, which possesses several health benefits, including anti-inflammatory, antioxidant, and antimicrobial properties, attributed to bioactive compounds, such as phenolics, carotenoids, and phytosterols<sup>146-148</sup>.

##### 4.12.1. Anti-inflammatory effects

*Phoenix dactylifera* L. exhibited significant anti-inflammatory activity. An Algerian date extract (50 mg/kg) reduced paw edema and inflammatory markers, such as C-Reactive Protein (CRP) and homocysteine, in mice<sup>149</sup>. Furthermore, methanolic extracts from Moroccan date seeds and fruits demonstrated potent effects in multiple models, reducing paw and ear edema in rodents. This activity is directly linked to their high phenolic and flavonoid content, with specific compounds, including rutin, quercetin, and caffeic acid, identified as key contributors<sup>150,151</sup>.

##### 4.12.2. Antioxidant activities

*Phoenix dactylifera* L. demonstrated notable and varied antioxidant potential. A hydroethanolic extract protected against oxidative stress by activating antioxidants, reducing ROS, and suppressing pro-apoptotic genes (p53, Bax)<sup>152</sup>. However, antioxidant capacity varies considerably among different date fruit and seed varieties, as shown by differences in polyphenol, flavonoid, and tannin content and their performance in FRAP, DPPH, and ABTS assays<sup>153,154</sup>. This highlights the need to evaluate each variety individually for specific applications.

##### 4.12.3. Immunomodulatory effects

*Phoenix dactylifera* L. exhibited potent antioxidant, anti-inflammatory, and immunomodulatory properties. In a rat



model of aluminum chloride-induced toxicity, a date fruit extract (500 mg/kg) effectively reversed oxidative stress by increasing glutathione, SOD, and catalase levels, while also decreasing pro-inflammatory cytokines (TNF- $\alpha$ , IL-6), NF- $\kappa$ B, and lipid markers peroxidation<sup>155</sup>.

#### 4.12.4. Treatment of lung injury

*Phoenix dactylifera* L. demonstrated significant protective effects against chemical-induced lung injury. In rats, an ethanolic extract of Ajwa date pulp mitigated benzo(a)pyrene-induced damage by restoring antioxidant enzymes (SOD, catalase, GPx) and reducing pro-inflammatory cytokines (TNF- $\alpha$ , IL-6, IFN- $\gamma$ ), CRP, and angiogenesis markers<sup>156</sup>. Similarly, date palm sap protected against bleomycin-induced lung fibrosis in rats by reducing lipid peroxidation, modulating hydroxyproline and antioxidant enzymes, and preventing morphological lesions, an effect attributed to its phenolic and vitamin content<sup>157</sup>.

#### 4.13. *Teucrium polium* L.

*Teucrium polium* L., a wild plant from the *Lamiaceae* family, is used in traditional medicine for its diverse therapeutic properties. These include treating diabetes, gastrointestinal disorders, inflammation, and rheumatism<sup>158,159</sup>. Its bioactivity is attributed to compounds, such as flavonoids, terpenoids, sterols, and saponins<sup>160</sup>.

##### 4.13.1. Anti-inflammatory effects

*Teucrium polium* L. exhibits significant anti-inflammatory activity through multiple mechanisms. Its hydroalcoholic and aqueous extracts reduce key pro-inflammatory cytokines (IL-6, TNF- $\alpha$ , IL-1 $\beta$ ) and markers (CRP, MCP-1) in rodent models of inflammation<sup>161-163</sup>. One study also indicated that can increase the anti-inflammatory cytokine IL-10<sup>163</sup>. These effects are consistently attributed to its polyphenol and flavonoid content<sup>161, 162</sup>.

##### 4.13.2. Antioxidant activities

*Teucrium polium* L. is a rich source of natural antioxidants. Studies consistently show that its methanolic extract possesses the highest antioxidant activity, correlating with its superior phenolic and flavonoid content<sup>164</sup>. Key antioxidant compounds identified include luteolin glycosides and pelargonin<sup>165</sup>. The plant's efficacy has been confirmed across multiple assays (DPPH, ABTS, FRAP,  $\beta$ -carotene bleaching), demonstrating significant free radical scavenging and reducing power<sup>164-166</sup>.

##### 4.13.3. Immunomodulatory effects

*Teucrium polium* L. exhibited selective immunomodulatory and cytotoxic effects. A methanolic extract promoted the proliferation of healthy human immune cells, increasing CD14+, CD3+, and CD20+ subsets with CD25+ activation markers, while inducing apoptosis in hepatitis C virus (HCV)-infected cells. This immunostimulatory effect was consistent in healthy cells but was only observed in infected cells at the highest extract concentration, demonstrating its potential as a targeted immunomodulatory agent<sup>167</sup>.

## 5. Conclusion

The present study validated Avicenna's traditional use of medicinal plants for tuberculosis (TB) by aligning it with modern scientific evidence. The plants exhibit multi-targeted therapeutic potential through four key mechanisms, including direct antimycobacterial activity, anti-inflammatory effects by suppressing TNF- $\alpha$ , IL-6, and IL-1 $\beta$ , potent antioxidant action, and immunomodulation. These properties support their role as promising adjunctive therapies to conventional TB treatment. While the findings are compelling, further *in vivo* and clinical research is necessary to fully determine their efficacy, safety, and best integration into modern anti-TB regimens for better patient outcomes.

## Declarations

### Competing interests

The authors declared that there is no conflict of interest.

### Authors' contributions

Zakiyeh Sakhavat Nia wrote the draft of the manuscript. Mehdi Sobhani and Zahra Sobhani revised the manuscript draft and reviewed the final edition of the article. All authors have reviewed and approved the final edition of the manuscript before publication in this journal.

### Ethical considerations

The authors declare that the present manuscript is original and is not being considered elsewhere for publication. The authors have reviewed additional ethical concerns, such as research misconduct, data fabrication, and redundancy.

### Funding

The authors declare that no funds, grants, or other support were received.

### Availability of data and materials

All data from the current study are available upon reasonable requests from the authors.

### Acknowledgments

The authors would like to thank the Research Council of Mashhad University of Medical Sciences, Iran, for financially supporting this study.

## References

1. Vasiliu A, Martinez L, Gupta RK, Hamada Y, Ness T, Kay A, et al. Tuberculosis prevention: current strategies and future directions. *Clin Microbiol Infect.* 2024; 30(9): 1123-1130. DOI: [10.1016/j.cmi.2023.10.023](https://doi.org/10.1016/j.cmi.2023.10.023)
2. Saelens JW, Sweeney MI, Viswanathan G, Xet-Mull AM, Smith KLJ, Sisk DM, et al. An ancestral mycobacterial effector promotes dissemination of infection. *Cell.* 2022; 185(24): 4507-4525. DOI: [10.1016/j.cell.2022.10.019](https://doi.org/10.1016/j.cell.2022.10.019)
3. Wei X, Yue L, Zhao B, Jiang N, Lei H, and Zhai X. Recent advances and challenges of revolutionizing drug-resistant tuberculosis treatment.

- Eur J Med Chem. 2024; 116785. DOI: [10.1016/j.ejmech.2024.116785](https://doi.org/10.1016/j.ejmech.2024.116785)
4. Rana HK, Singh AK, Kumar R, and Pandey AK. Antitubercular drugs: possible role of natural products acting as antituberculosis medication in overcoming drug resistance and drug-induced hepatotoxicity. *Naunyn Schmiedeberg's Arch Pharmacol*. 2024; 397(3): 1251-1273. DOI: [10.1007/s00210-023-02679-z](https://doi.org/10.1007/s00210-023-02679-z)
5. Raj R, Tripathi AK, Saranya P, Pal RS, Singh K, Jain D, et al. A review of molecular investigations on traditional chinese medicinal plant-based therapies in multidrug-resistant tuberculosis. *Pharmacol Res Mod Chin Med*. 2024; 100521. DOI: [10.1016/j.prmcm.2024.100521](https://doi.org/10.1016/j.prmcm.2024.100521)
6. Sobhani Z, Reza Nami S, Ahmad Emami S, Sahebkar A, and Javadi B. Medicinal plants targeting cardiovascular diseases in view of Avicenna. *Curr Pharm Des*. 2017; 23(17): 2428-2443. DOI: [10.2174/1381612823666170215104101](https://doi.org/10.2174/1381612823666170215104101)
7. Ghaffari F, Taheri M, Meyari A, Karimi Y, and Naseri M. Avicenna and clinical experiences in Canon of Medicine. *J Med Life*. 2022; 15(2): 168-178. DOI: [10.25122/jml-2021-0246](https://doi.org/10.25122/jml-2021-0246)
8. Liebenberg D, Gordhan BG, and Kana BD. Drug resistant tuberculosis: Implications for transmission, diagnosis, and disease management. *Front Cell Infect Microbiol*. 2022; 23: 12: 943545. DOI: [10.3389/fcimb.2022.943545](https://doi.org/10.3389/fcimb.2022.943545)
9. Kayongo A, Nyiro B, Siddharthan T, Kirenga B, Checkley W, Lutaakome Joloba M, et al. Mechanisms of lung damage in tuberculosis: implications for chronic obstructive pulmonary disease. *Front Cell Infect Microbiol*. 2023; 13: 1146571. DOI: [10.3389/fcimb.2023.1146571](https://doi.org/10.3389/fcimb.2023.1146571)
10. World health organization (WHO). World health organization global tuberculosis report 2024. Geneva: World Health Organization; 2024. Available at: <https://iris.who.int/server/api/core/bitstreams/7292c91e-ffb0-4cef-ac39-0200f06961ea/content>
11. Cadena AM, Fortune SM, and Flynn JL. Heterogeneity in tuberculosis. *Nat Rev Immunol*. 2017; 17(11): 691-702. DOI: [10.1038/nri.2017.69](https://doi.org/10.1038/nri.2017.69)
12. Chee CBE, Reves R, Zhang Y, and Belknap R. Latent tuberculosis infection: Opportunities and challenges. *Respirology*. 2018; 23(10): 893-900. DOI: [10.1111/resp.13346](https://doi.org/10.1111/resp.13346)
13. Natarajan A, Beena PM, Devnikar AV, and Mali S. A systemic review on tuberculosis. *Indian J Tuberc*. 2020; 67(3): 295-311. DOI: [10.1016/j.ijtb.2020.02.005](https://doi.org/10.1016/j.ijtb.2020.02.005)
14. Chai Q, Lu Z, and Liu CH. Host defense mechanisms against *Mycobacterium tuberculosis*. *Cell Mol Life Sci*. 2020; 77(10): 1859-1878. DOI: [10.1007/s00018-019-03353-5](https://doi.org/10.1007/s00018-019-03353-5)
15. de Martino M, Lodi L, Galli L, and Chiappini E. Immune response to *Mycobacterium tuberculosis*: A narrative review. *Front Pediatr*. 2019; 7: 350. DOI: [10.3389/fped.2019.00350](https://doi.org/10.3389/fped.2019.00350)
16. Alarcón V, Alarcón E, Figueroa C, and Mendoza-Ticona A. Tuberculosis in Peru: epidemiological situation, progress and challenges for its control. *Rev Peru Med Exp Salud Publica*. 2017; 34(2): 299-310. DOI: [10.17843/rpmesp.2017.342.2384](https://doi.org/10.17843/rpmesp.2017.342.2384)
17. Flynn JL and Chan J. Immune cell interactions in tuberculosis. *Cell*. 2022; 185(25): 4682-4702. DOI: [10.1016/j.cell.2022.10.025](https://doi.org/10.1016/j.cell.2022.10.025)
18. Hamada Y, Getahun H, Tadesse BT, and Ford N. HIV-associated tuberculosis. *Int J STD AIDS*. 2021; 32(9): 780-790. DOI: [10.1177/0956462421992257](https://doi.org/10.1177/0956462421992257)
19. Ferraris DM, Miggiano R, Rossi F, and Rizzi M. *Mycobacterium tuberculosis* molecular determinants of infection, survival strategies, and vulnerable targets. *Pathogens*. 2018; 7(1): 17. DOI: [10.3390/pathogens7010017](https://doi.org/10.3390/pathogens7010017)
20. Goossens SN, Sampson SL, and Van Rie A. Mechanisms of drug-induced tolerance in *Mycobacterium tuberculosis*. *Clin Microbiol Rev*. 2020; 34(1): e00141-20. DOI: [10.1128/CMR.00141-20](https://doi.org/10.1128/CMR.00141-20)
21. Godfrey MS and Friedman LN. Tuberculosis and biologic therapies: Anti-tumor necrosis Factor- $\alpha$  and beyond. *Clin Chest Med*. 2019; 40(4): 721-739. DOI: [10.1016/j.ccm.2019.07.003](https://doi.org/10.1016/j.ccm.2019.07.003)
22. Khader SA, Divangahi M, Hanekom W, Hill PC, Maeurer M, Makar KW, et al. Targeting innate immunity for tuberculosis vaccination. *J Clin Invest*. 2020; 129(9): 3482-3491. DOI: [10.1172/JCI128877](https://doi.org/10.1172/JCI128877)
23. Chai Q, Wang L, Liu CH, and Ge B. New insights into the evasion of host innate immunity by *Mycobacterium tuberculosis*. *Cell Mol Immunol*. 2020; 17(9): 901-913. DOI: [10.1038/s41423-020-0502-z](https://doi.org/10.1038/s41423-020-0502-z)
24. Sinigaglia A, Peta E, Riccetti S, Venkateswaran S, Manganelli R, and Barzon L. Tuberculosis-associated microRNAs: From pathogenesis to disease biomarkers. *Cells*. 2020; 9(10): 2160. DOI: [10.3390/cells9102160](https://doi.org/10.3390/cells9102160)
25. Sina I. Al-Qanun fi al-Tibb. 1st ed. Beirut: Dar al-Kutub al-Ilmiyah; 2005.
26. Abdollahinia A, Naseri M, Tahmasbi S, Adimi P, Sadr M, and Velayati AA. Ideal lifestyle to have healthy lungs: Persian medicine viewpoint. *Tradit Integr Med*. 2022; 7(1): 150-158. DOI: [10.18502/tim.v7i1.9071](https://doi.org/10.18502/tim.v7i1.9071)
27. Szopa A, Pajor J, Klin P, Rzepiela A, Elansary HO, Al-Mana FA, et al. *Artemisia absinthium* L.-Importance in the history of medicine, the latest advances in phytochemistry and therapeutical, cosmetological and culinary uses. *Plants*. 2020; 9(9): 1063. DOI: [10.3390/plants9091063](https://doi.org/10.3390/plants9091063)
28. Hussain M, Raja NI, Akram A, Iftikhar A, Ashfaq D, Yasmeen F, et al. A status review on the pharmacological implications of *Artemisia absinthium*: A critically endangered plant. *Asian Pac J Trop Dis*. 2017; 7(3): 185-192. DOI: [10.12980/apjtd.7.2017D6-385](https://doi.org/10.12980/apjtd.7.2017D6-385)
29. Nigam M, Atanassova M, Mishra AP, Pezzani R, Devkota HP, Plygun S, et al. Bioactive compounds and health benefits of *Artemisia* species. *Nat Prod Commun*. 2019; 14(7): 1934578X19850354. DOI: [10.1177/1934578X19850354](https://doi.org/10.1177/1934578X19850354)
30. Anibogwu R, Jesus KD, Pradhan S, Pashikanti S, Mateen S, and Sharma K. Extraction, isolation and characterization of bioactive compounds from *Artemisia* and their biological significance: A review. *Molecules*. 2021; 26(22): 6995. DOI: [10.3390/molecules26226995](https://doi.org/10.3390/molecules26226995)
31. Batiha GE, Olatunde A, El-Mleeh A, Hetta HF, Al-Rejaie S, Alghamdi S, et al. Bioactive compounds, pharmacological actions, and pharmacokinetics of Wormwood (*Artemisia absinthium*). *Antibiotics*. 2020; 9(6): 353. DOI: [10.3390/antibiotics9060353](https://doi.org/10.3390/antibiotics9060353)
32. Amrollahi H, Nazari H, Parvini S, Nazari N, and Mohammadi A. Anti-inflammatory and analgesic activities of *Artemisia absinthium* and chemical composition of its essential oil. *Int J Pharm Sci Rev Res*. 2014; 38: 237-244.
33. Neagu E, Paun G, Albu C, Apreutesei OT, and Radu GL. *In vitro* assessment of the antidiabetic and anti-inflammatory potential of *Artemisia absinthium*, *Artemisia vulgaris* and *Trigonella foenum-graecum* extracts processed using membrane technologies. *Molecules*. 2023; 28(20): 7156. DOI: [10.3390/molecules28207156](https://doi.org/10.3390/molecules28207156)
34. Craciunescu O, Constantin D, Gaspar A, Toma L, Utoiu E, and Moldovan L. Evaluation of antioxidant and cytoprotective activities of *Arnica montana* L. and *Artemisia absinthium* L. ethanolic extracts. *Chem Cent J*. 2012; 6: 97. DOI: [10.1186/1752-153X-6-97](https://doi.org/10.1186/1752-153X-6-97)
35. Amat N, Upur H, and Blažeković B. *In vivo* hepatoprotective activity of the aqueous extract of *Artemisia absinthium* L. against chemically and immunologically induced liver injuries in mice. *J Ethnopharmacol*. 2010; 131(2): 478-484. DOI: [10.1016/j.jep.2010.07.023](https://doi.org/10.1016/j.jep.2010.07.023)
36. Ikram M, Shafi N, Mir I, Do M, Nguyen P, and Le Quesne P. 24 $\beta$ -Ethylcholesta-7, 22-Dien-3 $\beta$ -ol: A possibly antipyretic constituent of *Artemisia absinthium*. *Planta Med*. 1987; 53(04): 389-389. DOI: [10.1055/s-2006-962748](https://doi.org/10.1055/s-2006-962748)
37. Bhat MM, Ansari AP, Ahmad A, Qayoom I, and Reshi BM. Antipyretic activity of the hydro-alcoholic extract of *Artemisia absinthium* L. as a stand-alone and as an adjuvant with barley water against yeast-induced pyrexia in albino Wistar rats. *J Complement Integr Med*. 2023. DOI: [10.1515/jcim-2023-0307](https://doi.org/10.1515/jcim-2023-0307)
38. Hojageldiyev T, Bolmammedov Y, and Gurbanaliyev S. Antimycobacterial activity of ethanolic extract of *Artemisia absinthium* L. *World Sci News*. 2019; 119: 224-230.
39. Lee JK. Anti-inflammatory effects of eriodictyol in lipopolysaccharide-stimulated raw 264.7 murine macrophages. *Arch Pharm Res*. 2011; 34(4): 671-679. DOI: [10.1007/s12272-011-0418-3](https://doi.org/10.1007/s12272-011-0418-3)
40. Abiri R, Silva ALM, de Mesquita LSS, de Mesquita JWC, Atabaki N, de Almeida EB, et al. Towards a better understanding of *Artemisia vulgaris*: Botany, phytochemistry, pharmacological and biotechnological potential. *Food Res Int*. 2018; 109: 403-415. DOI: [10.1016/j.foodres.2018.03.072](https://doi.org/10.1016/j.foodres.2018.03.072)
41. Trifan A, Zengin G, Sinan KI, Sieniawska E, Sawicki R, Maciejewska-Turska M, et al. Unveiling the phytochemical profile and biological potential of five *Artemisia* species. *Antioxidants*. 2022; 11(5): 1017. DOI: [10.3390/antiox11051017](https://doi.org/10.3390/antiox11051017)
42. Soon L, Ng PQ, Chellian J, Madheswaran T, Panneerselvam J, Gupta G, et al. Therapeutic potential of *Artemisia vulgaris*: An insight into underlying immunological mechanisms. *J Environ Pathol Toxicol Oncol*. 2019; 38(3): 205-216. DOI: [10.1615/JEnvironPatholToxicolOncol.2019029397](https://doi.org/10.1615/JEnvironPatholToxicolOncol.2019029397)
43. Pandey J, Bhusal S, Nepali L, Khatrri M, Ramdam R, Barakoti H, et al. Anti-inflammatory activity of *Artemisia vulgaris* leaves, originating from three different altitudes of Nepal. *Sci World J*. 2021; 2021: 6678059.

- DOI: [10.1155/2021/6678059](https://doi.org/10.1155/2021/6678059)
44. Ashok PK and Upadhyaya K. Evaluation of analgesic and anti-inflammatory activities of aerial parts of *Artemisia vulgaris* L. in experimental animal models. J Biol Act Prod Nat. 2013; 3(1): 101-105. DOI: [10.1080/22311866.2013.782761](https://doi.org/10.1080/22311866.2013.782761)
  45. Temraz A and El-Tantawy WH. Characterization of antioxidant activity of extract from *Artemisia vulgaris*. Pak J Pharm Sci. 2008; 21(4): 321-326.
  46. Ben Nasr S, Aazza S, Mnif W, and Miguel M. In-vitro antioxidant and anti-inflammatory activities of *Pituranthos chloranthus* and *Artemisia vulgaris* from Tunisia. Int J Appl Pharm Sci Res. 2020; 11(2): 605-614.
  47. Khan AU and Gilani AH. Antispasmodic and bronchodilator activities of *Artemisia vulgaris* are mediated through dual blockade of muscarinic receptors and calcium influx. J Ethnopharmacol. 2009; 126(3): 480-486. DOI: [10.1016/j.jep.2009.09.010](https://doi.org/10.1016/j.jep.2009.09.010)
  48. Lim TK. *Glycyrrhiza glabra*. Edible Med Non-Med Plants. 2015; 22: 354-457. DOI: [10.1007/978-94-017-7276-1\\_18](https://doi.org/10.1007/978-94-017-7276-1_18)
  49. Zang Y. Pharmacological activities of coumarin compounds in Licorice: A review. Nat Prod Commun. 2020; 15(9): 1934578X20953954. DOI: [10.1177/1934578X20953954](https://doi.org/10.1177/1934578X20953954)
  50. Sun J, Zhang Q, Yang G, Li Y, Fu Y, Zheng Y, et al. The licorice flavonoid isoliquiritigenin attenuates *Mycobacterium tuberculosis*-induced inflammation through Notch1/NF- $\kappa$ B and MAPK signaling pathways. J Ethnopharmacol. 2022; 294: 115368. DOI: [10.1016/j.jep.2022.115368](https://doi.org/10.1016/j.jep.2022.115368)
  51. Frattaruolo L, Carullo G, Brindisi M, Mazzotta S, Bellissimo L, Rago V, et al. Antioxidant and anti-inflammatory activities of flavanones from *Glycyrrhiza glabra* L. (licorice) leaf phytocomplexes: Identification of licoflavanone as a modulator of NF- $\kappa$ B/MAPK pathway. Antioxidants. 2019; 8(6): 186. DOI: [10.3390/antiox8060186](https://doi.org/10.3390/antiox8060186)
  52. Ma C, Ma Z, Liao XL, Liu J, Fu Q, and Ma S. Immunoregulatory effects of glycyrrhizic acid exerts anti-asthmatic effects via modulation of Th1/Th2 cytokines and enhancement of CD4+ CD25+ Foxp3+ regulatory T cells in ovalbumin-sensitized mice. J Ethnopharmacol. 2013; 148(3): 755-762. DOI: [10.1016/j.jep.2013.04.021](https://doi.org/10.1016/j.jep.2013.04.021)
  53. Wu Q, Tang Y, Zhang J, Hu X, Wang Q, and Huang J. Therapeutic effects of glycyrrhizic acid on asthma airway inflammation in mice and its mechanism. Zhonghua Yi Xue Za Zhi. 2014; 94(42): 3338-3344.
  54. Shitole M and Pawar V. Study of potential antitussive activity of *Glycyrrhiza glabra* granules by using a cough model induced by Sulphur dioxide gas in mice. Asian J Pharm Clin Res. 2019; 12(10): 262-267. DOI: [10.22159/ajpcr.2019.v12i10.33967](https://doi.org/10.22159/ajpcr.2019.v12i10.33967)
  55. Kuang Y, Li B, Fan J, Qiao X, and Ye M. Antitussive and expectorant activities of licorice and its major compounds. Bioorg Med Chem. 2018; 26(1): 278-284. DOI: [10.1016/j.bmc.2017.11.046](https://doi.org/10.1016/j.bmc.2017.11.046)
  56. Saha S, Nosál'ová G, Ghosh D, Flešková D, Capek P, and Ray B. Structural features and *in vivo* antitussive activity of the water extracted polymer from *Glycyrrhiza glabra*. Int J Biol Macromol. 2011; 48(4): 634-638. DOI: [10.1016/j.ijbiomac.2011.02.003](https://doi.org/10.1016/j.ijbiomac.2011.02.003)
  57. Zadeh JB, Kor ZM, and Gofgar MM. Licorice (*Glycyrrhiza glabra* Linn) as a valuable medicinal plant. Int J Adv Biol Biomed Res. 2013; 1(10): 1281-1288.
  58. Martins N, Barros L, Dueñas M, Santos-Buelga C, and Ferreira IC. Characterization of phenolic compounds and antioxidant properties of *Glycyrrhiza glabra* L. rhizomes and roots. RSC Adv. 2015; 5(34): 26991-26997. DOI: [10.1039/C5RA03963K](https://doi.org/10.1039/C5RA03963K)
  59. Sharma V and Agrawal R. *In vivo* antioxidant and hepatoprotective potential of *Glycyrrhiza glabra* extract on carbon tetra chloride (CCl<sub>4</sub>) induced oxidative-stress mediated hepatotoxicity. Int J Res Med Sci. 2014; 2(1): 314-320. DOI: [10.5455/2320-6012.ijrms20140260](https://doi.org/10.5455/2320-6012.ijrms20140260)
  60. Shang H, Cao S, Wang J, Zheng H, and Putheti R. Glabridin from Chinese herb licorice inhibits fatigue in mice. Afr J Tradit Complement Altern Med. 2010; 7(1): 17-23. DOI: [10.4314/ajtcam.v7i1.57225](https://doi.org/10.4314/ajtcam.v7i1.57225)
  61. Trivedi R and Sharma K. Hydroalcoholic extract of *Glycyrrhiza glabra* Linn. attenuates chronic fatigue stress induced behavioral alterations in mice. Int J Pharm Biol Arch. 2011; 2(3): 996-1001.
  62. Am Lee S, Lee SH, Kim JY, and Lee WS. Effects of glycyrrhizin on lipopolysaccharide-induced acute lung injury in a mouse model. J Thorac Dis. 2019; 11(4): 1287-1302. DOI: [10.21037/jtd.2019.04.14](https://doi.org/10.21037/jtd.2019.04.14)
  63. Wang J, Ren C, Bi W, and Batu W. Glycyrrhizin mitigates acute lung injury by inhibiting the NLRP3 inflammasome *in vitro* and *in vivo*. J Ethnopharmacol. 2023; 303: 115948. DOI: [10.1016/j.jep.2022.115948](https://doi.org/10.1016/j.jep.2022.115948)
  64. Viswanathan V, Pharande R, Bannalika A, Gupta P, Gupta U, and Mukhe A. Inhalable liposomes of *Glycyrrhiza glabra* extract for use in tuberculosis: formulation, *in vitro* characterization, *in vivo* lung deposition, and *in vivo* pharmacodynamic studies. Drug Dev Ind Pharm. 2019; 45(1): 11-20. DOI: [10.1080/03639045.2018.1513025](https://doi.org/10.1080/03639045.2018.1513025)
  65. Grover IS, Rai J, Kajal NC, and Bhushan B. Effect of liquorice [*Glycyrrhiza glabra* Linn.] As an adjuvant in newly diagnosed sputum smear-positive patients of pulmonary tuberculosis on directly observed treatment short course (dots) therapy. Chest. 2006; 130(4): 95S. DOI: [10.1378/chest.130.4.MeetingAbstracts.95S-c](https://doi.org/10.1378/chest.130.4.MeetingAbstracts.95S-c)
  66. Kumar V, Kaur N, Kaur A, and Wadhwa P. Phytochemistry and pharmacology of Indian traditional plant hyssop (*Hyssopus officinalis* L.): A Review. Nat Prod J. 2023; 13(4): 11-41. DOI: [10.2174/2210315512666220811153919](https://doi.org/10.2174/2210315512666220811153919)
  67. Hanganu D, Pârnu A, Mărculescu A, Oniga I, Tipericiu B, and Benedec D. *Hyssopus officinalis* L. (fam. Lamiaceae) a potential plant food supplements with anti-inflammatory effect. J Eco Agri Tourism. 2016; 12(2): 10-14.
  68. Mićović T, Stanković JSK, Bauer R, Nöst X, Marković Z, Milenković D, et al. *In vitro*, *in vivo* and *in silico* evaluation of the anti-inflammatory potential of *Hyssopus officinalis* L. subsp. aristatus (Godr.) Nyman (Lamiaceae). J Ethnopharmacol. 2022; 293: 115201. DOI: [10.1016/j.jep.2022.115201](https://doi.org/10.1016/j.jep.2022.115201)
  69. Ma X, Ma X, Ma Z, Wang J, Sun Z, Yu W, et al. Effect of *Hyssopus officinalis* L. on inhibiting airway inflammation and immune regulation in a chronic asthmatic mouse model. Exp Ther Med. 2014; 8(5): 1371-1374. DOI: [10.3892/etm.2014.1978](https://doi.org/10.3892/etm.2014.1978)
  70. Ma X, Ma X, Ma Z, Sun Z, Yu W, Wang J, et al. The effects of uygur herb *Hyssopus officinalis* L. on the process of airway remodeling in asthmatic mice. Evid Based Complement Alternat Med. 2014; 2014: 710870. DOI: [10.1155/2014/710870](https://doi.org/10.1155/2014/710870)
  71. Alinezhad H, Azimi R, Zare M, Ebrahimzadeh MA, Eslami S, Nabavi SF, et al. Antioxidant and antihemolytic activities of ethanolic extract of flowers, leaves, and stems of *Hyssopus officinalis* L. Var. angustifolius. Int J Food Prop. 2013; 16(5): 1169-1178. DOI: [10.1080/10942912.2011.578319](https://doi.org/10.1080/10942912.2011.578319)
  72. Özer H, Sökmen M, Güllüce M, Adigüzel A, Kilic H, Şahin F, et al. *In vitro* antimicrobial and antioxidant activities of the essential oils and methanol extracts of *Hyssopus officinalis* L. ssp. *angustifolius*. Ital J Food Sci. 2006; 18(1): 33-48.
  73. Fathiazad F, Mazandarani M, and Hamedeyazdan S. Phytochemical analysis and antioxidant activity of *Hyssopus officinalis* L. from Iran. Adv Pharm Bull. 2011; 1(2): 63-67.
  74. Henna A, Nemmiche S, Dandlen S, and Miguel MG. *Myrtus communis* essential oils: Insecticidal, antioxidant and antimicrobial activities: A review. J Essent Oil Res. 2019; 31(6): 487-545. DOI: [10.1080/10412905.2019.1611672](https://doi.org/10.1080/10412905.2019.1611672)
  75. Akbar S and Akbar S. *Myrtus communis* L. (Myrtaceae). Handbook of 200 Medicinal Plants: A comprehensive review of their traditional medical uses and scientific justifications. 2020: 1251-1262. DOI: [10.1007/978-3-030-16807-0\\_131](https://doi.org/10.1007/978-3-030-16807-0_131)
  76. Dabbaghi MM, Fadaei MS, Soleimani Roudi H, Baradaran Rahimi V, and Askari VR. A review of the biological effects of *Myrtus communis*. Physiol Rep. 2023; 11(14): e15770. DOI: [10.14814/phy2.15770](https://doi.org/10.14814/phy2.15770)
  77. Khosropour P, Sajjadi S-E, Talebi A, and Minaian M. Anti-inflammatory effect of *Myrtus communis* hydroalcoholic extract and essential oil on acetic acid-induced colitis in rats. J Rep Pharm Sci. 2019; 8(2): 204-210. DOI: [10.4103/jrptps.JRPTPS\\_8\\_19](https://doi.org/10.4103/jrptps.JRPTPS_8_19)
  78. Feišt C, Franke L, Appendino G, and Werz O. Identification of molecular targets of the oligomeric nonprenylated acylphloroglucinols from *Myrtus communis* and their implication as anti-inflammatory compounds. J Pharmacol Exp Ther. 2005; 315(1): 389-396. DOI: [10.1124/jpet.105.090720](https://doi.org/10.1124/jpet.105.090720)
  79. Rossi A, Di Paola R, Mazzon E, Genovese T, Caminiti R, Bramanti P, et al. Myrtucommulone from *Myrtus communis* exhibits potent anti-inflammatory effectiveness *in vivo*. J Pharmacol Exp Ther. 2009; 329(1): 76-86. DOI: [10.1124/jpet.108.143214](https://doi.org/10.1124/jpet.108.143214)
  80. Wannes WA, Mhamdi B, Sriti J, Jemia MB, Ouchikh O, Hamdaoui G, et al. Antioxidant activities of the essential oils and methanol extracts from myrtle (*Myrtus communis* var. *italica* L.) leaf, stem and flower. Food Chem Toxicol. 2010; 48(5): 1362-1370. DOI: [10.1016/j.fct.2010.03.002](https://doi.org/10.1016/j.fct.2010.03.002)
  81. Amensour M, Sendra E, Abrini J, Bouhdid S, Pérez-Alvarez JA, and Fernández-López J. Total phenolic content and antioxidant activity of myrtle (*Myrtus communis*) extracts. Nat Prod Commun. 2009; 4(6): 1934578X0900400616. DOI: [10.1177/1934578X0900400616](https://doi.org/10.1177/1934578X0900400616)
  82. Taher MS, Salloom YF, Al-Asadi RA, Al-Mousswi ZJ, and Alamrani HA.



- The medicinal importance of Thyme plant (*Thymus vulgaris*). Biomedicine. 2021; 29;41(3):531-4. DOI: [10.51248/v4i13.708](https://doi.org/10.51248/v4i13.708)
83. Grespan R, Aguiar RP, Giubilei FN, Fuso RR, Damião MJ, Silva EL, et al. Hepatoprotective effect of pretreatment with *Thymus vulgaris* essential oil in experimental model of acetaminophen-induced injury. Evid Based Complement Alternat Med. 2014; 2014: 954136. DOI: [10.1155/2014/954136](https://doi.org/10.1155/2014/954136)
  84. Thompson JD, Chalchat JC, Michet A, Linhart YB, and Ehlers B. Qualitative and quantitative variation in monoterpene co-occurrence and composition in the essential oil of *Thymus vulgaris* chemotypes. J Chem Ecol. 2003; 29(4): 859-880. DOI: [10.1023/A:1022927615442](https://doi.org/10.1023/A:1022927615442)
  85. de Lira Mota KS, de Oliveira Pereira F, de Oliveira WA, Lima IO, and de Oliveira Lima E. Antifungal activity of *Thymus vulgaris* L. essential oil and its constituent phytochemicals against *Rhizopus oryzae*: Interaction with ergosterol. Molecules. 2012; 17(12): 14418-14433. DOI: [10.3390/molecules171214418](https://doi.org/10.3390/molecules171214418)
  86. Benameur Q, Gervasi T, Pellizzeri V, Pluchtová M, Tali-Maama H, Assaouf F, et al. Antibacterial activity of *Thymus vulgaris* essential oil alone and in combination with cefotaxime against blaESBL producing multidrug resistant *Enterobacteriaceae* isolates. Nat Prod Res. 2019; 33(18): 2647-2654. DOI: [10.1080/14786419.2018.1466124](https://doi.org/10.1080/14786419.2018.1466124)
  87. Fachini-Queiroz FC, Kummer R, Estevão-Silva CF, Carvalho MD, Cunha JM, Grespan R, et al. Effects of thymol and carvacrol, constituents of *Thymus vulgaris* L. essential oil, on the inflammatory response. Evid Based Complement Alternat Med. 2012; 2012: 657026. DOI: [10.1155/2012/657026](https://doi.org/10.1155/2012/657026)
  88. Ocaña A and Reglero G. Effects of thyme extract oils (from *Thymus vulgaris*, *Thymus zygis*, and *Thymus hyemalis*) on cytokine production and gene expression of oxLDL-stimulated THP-1-macrophages. J Obes. 2012; 2012: 104706. DOI: [10.1155/2012/104706](https://doi.org/10.1155/2012/104706)
  89. Mousa AM, Almatroudi A, Alwashmi AS, Al Abdulmonem W, Aljohani AS, Alhumaydhi FA, et al. Thyme oil alleviates Ova-induced bronchial asthma through modulating Th2 cytokines, IgE, TSLP and ROS. Biomed Pharmacother. 2021; 140: 111726. DOI: [10.1016/j.biopha.2021.111726](https://doi.org/10.1016/j.biopha.2021.111726)
  90. Al-Khalaf MI. Thyme and thymol effects on induced bronchial asthma in mice. Life Sci J. 2013; 10(2): 693-699. DOI: [10.7813/2075-4124.2013/5-2/A.25](https://doi.org/10.7813/2075-4124.2013/5-2/A.25)
  91. Aldosary S, El-Rahman S, Al-Jameel S, and Alromihi N. Antioxidant and antimicrobial activities of *Thymus vulgaris* essential oil contained and synthesis thymus (*Vulgaris*) silver nanoparticles. Braz J Biol. 2021; 83: e244675. DOI: [10.1590/1519-6984.244675](https://doi.org/10.1590/1519-6984.244675)
  92. Rahgozar N, Bakhshi Khaniki G, and Sardari S. Evaluation of antimycobacterial and synergistic activity of plants selected based on cheminformatic parameters. Iran Biomed J. 2018; 22(6): 401-407. DOI: [10.29252/22.6.401](https://doi.org/10.29252/22.6.401)
  93. Lall N and Meyer JJ. *In vitro* inhibition of drug-resistant and drug-sensitive strains of *Mycobacterium tuberculosis* by ethnobotanically selected South African plants. J Ethnopharmacol. 1999; 66(3): 347-354. DOI: [10.1016/S0378-8741\(98\)00185-8](https://doi.org/10.1016/S0378-8741(98)00185-8)
  94. Dizaji P. *In vitro* antibacterial activity of *Thymus vulgaris* essential oil against *Mycobacterium tuberculosis*. Infect Epidemiol Microbiol. 2018; 4(2): 47-51.
  95. Ramazanadeh R, Marzban A, and Shakib P. Anti-*Mycobacterium tuberculosis* effects of folk medicinal plants in Iran: A Mini-systematic review. Iran J Med Microbiol. 2023; 17(1): 1-13. DOI: [10.30699/ijmm.17.1.1](https://doi.org/10.30699/ijmm.17.1.1)
  96. Boskabady MH, Shafei MN, Saberi Z, and Amini S. Pharmacological effects of *Rosa damascena*. Iran J Basic Med Sci. 2011; 14(4): 295-307.
  97. Fatemi F, Golbodagh A, Hojhosseini R, Dadkhah A, Akbarzadeh K, Salome D, et al. Anti-inflammatory effects of deuterium-depleted water plus *rosa damascena* mill. Essential oil via cyclooxygenase-2 pathway in rats. Turk J Pharm Sci. 2020; 17(1): 99-106. DOI: [10.4274/tjps.galenos.2018.24381](https://doi.org/10.4274/tjps.galenos.2018.24381)
  98. Dadkhah A, Fatemi F, Mohammadi Malayeri MR, Karvin Ashtiani MH, Mosavi Z, Najj S, et al. The anti-inflammatory and antioxidant effects of *Rosa damascena* Mill. essential oil on the lung injury in the CLP model. J Med Plants. 2020; 19(74): 277-294. DOI: [10.29252/jmp.19.74.277](https://doi.org/10.29252/jmp.19.74.277)
  99. Chroho M, Bouymajane A, Oulad El Majdoub Y, Cacciola F, Mondello L, Aazza M, et al. Phenolic composition, antioxidant and antibacterial activities of extract from flowers of *Rosa damascena* from Morocco. Separations. 2022; 9(9): 247. DOI: [10.3390/separations9090247](https://doi.org/10.3390/separations9090247)
  100. Alizadeh Z and Fattahi M. Essential oil, total phenolic, flavonoids, anthocyanins, carotenoids and antioxidant activity of cultivated Damask Rose (*Rosa damascena*) from Iran: With chemotyping approach concerning morphology and composition. Sci Hortic. 2021; 288: 110341. DOI: [10.1016/j.scienta.2021.110341](https://doi.org/10.1016/j.scienta.2021.110341)
  101. Mawarni E, Ginting CN, Chiuman L, Girsang E, Handayani RAS, and Widowati W. Antioxidant and elastase inhibitor potential of petals and receptacle of rose flower (*Rosa damascena*). Pharm Sci Res. 2020; 7(4): 1-8. DOI: [10.7454/psr.v7i2.1016](https://doi.org/10.7454/psr.v7i2.1016)
  102. Boskabady M, Kiani S, and Rakhshandah H. Relaxant effects of *Rosa damascena* on guinea pig tracheal chains and its possible mechanism(s). J Ethnopharmacol. 2006; 106(3): 377-382. DOI: [10.1016/j.jep.2006.01.013](https://doi.org/10.1016/j.jep.2006.01.013)
  103. Demirel S. *Rosa damascena* Miller essential oil relaxes rat trachea via KV channels, KATP channels, and BKCa channels. Prostaglandins Other Lipid Mediat. 2022; 163: 106673. DOI: [10.1016/j.prostaglandins.2022.106673](https://doi.org/10.1016/j.prostaglandins.2022.106673)
  104. Raghuvanshi D, Dhalaria R, Sharma A, Kumar D, Kumar H, Valis M, et al. Ethnomedicinal plants traditionally used for the treatment of Jaundice (Icterus) in Himachal Pradesh in Western Himalaya-A review. Plants. 2021; 10(2): 232. DOI: [10.3390/plants10020232](https://doi.org/10.3390/plants10020232)
  105. Singh M, Singh N, Khare PB, and Rawat AK. Antimicrobial activity of some important *Adiantum* species used traditionally in indigenous systems of medicine. J Ethnopharmacol. 2008; 115(2): 327-329. DOI: [10.1016/j.jep.2007.09.018](https://doi.org/10.1016/j.jep.2007.09.018)
  106. Dehdari S and Hajimehdipoor H. Medicinal properties of *Adiantum capillus-veneris* Linn. in traditional medicine and modern phytotherapy: A review article. Iran J Public Health. 2018; 47(2): 188-197.
  107. Khoramian L, Sajjadi S-E, and Minaian M. Anti-inflammatory effect of *Adiantum capillus-veneris* hydroalcoholic and aqueous extracts on acetic acid-induced colitis in rats. Avicenna J Phytomed. 2020; 10(5): 492-503.
  108. Ullah S, Jan G, Gul F, Khan S, Khattak M, Bibi H, et al. Phytochemistry, anti-inflammatory and antipyretic activities of *Adiantum capillus-veneris* in Swiss albino mice. Int J Fauna Biol Stud. 2018; 5(3): 19-25.
  109. Seif M, Aati H, Amer M, Ragauskas AJ, Seif A, El-Sappah AH, et al. Mitigation of hepatotoxicity via boosting antioxidants and reducing oxidative stress and inflammation in carbendazim-treated rats using *Adiantum Capillus-Veneris* L. Extract. Molecules. 2023; 28(12): 4720. DOI: [10.3390/molecules28124720](https://doi.org/10.3390/molecules28124720)
  110. Khodaie L, Esnaashari S, and Moghaddam SB. Essential oil of arial parts of *Adiantum capillus-veneris*: Chemical composition and antioxidant activity. Jundishapur J Nat Pharm Prod. 2015; 10(4): e21968. DOI: [10.17795/jjnpp-21968](https://doi.org/10.17795/jjnpp-21968)
  111. Abdulqadir A, Cakmak YS, and Zengin G. Phenolic compounds, antioxidant properties and enzyme inhibition ability of *Adiantum capillus veneris* L. linked to alzheimer's disease, diabetes mellitus and skin disorders. Curr Org Chem. 2018; 22(17): 1697-1703. DOI: [10.2174/1385272822666180711145256](https://doi.org/10.2174/1385272822666180711145256)
  112. Boukada F, Sitayeb S, Khadem H, Meddah B, and Zohra SF. Chemical composition, antioxidant and antibacterial activity of *Adiantum capillus-veneris* L. extract from Algeria. Kragujevac J Sci. 2022; 44: 91-101. DOI: [10.5937/KgJSci2244091B](https://doi.org/10.5937/KgJSci2244091B)
  113. Yadegari M, Riahy S, Mirdar S, Hamidian G, Afkhami SM, Saeidi A, et al. The TNF- $\alpha$ , P53 protein response and lung respiratory changes related to exercise, chronic hypoxia and *adiantum capillus-veneris* supplementation. Adv Respir Med. 2019; 87(4): 226-234. DOI: [10.5603/ARM.2019.0037](https://doi.org/10.5603/ARM.2019.0037)
  114. Yadegari M, Sellami M, Riahy S, Mirdar S, Hamidian G, Saeidi A, et al. Supplementation of *Adiantum capillus-veneris* modulates alveolar apoptosis under hypoxia condition in Wistar rats exposed to exercise. Medicina. 2019; 55(7): 401. DOI: [10.3390/medicina55070401](https://doi.org/10.3390/medicina55070401)
  115. Piri F, Mirdar S, and Hedayati M. The interactive effect of interval training and ethanol extract of *Adiantum Capillus Veneris* on the levels of metallothionein in lung male rats. Complement Med J. 2019; 8(4): 3467-3477.
  116. Akram M. Minireview on *Achillea millefolium* Linn. J Membr Biol. 2013; 246(9): 661-663. DOI: [10.1007/s00232-013-9588-x](https://doi.org/10.1007/s00232-013-9588-x)
  117. Ali SI, Gopalakrishnan B, and Venkatesalu V. Pharmacognosy, phytochemistry and pharmacological properties of *Achillea millefolium* L.: A review. Phytother Res. 2017; 31(8): 1140-1161. DOI: [10.1002/ptr.5840](https://doi.org/10.1002/ptr.5840)
  118. Ngo HT, Hwang E, Kang H, Park B, Seo SA, and Yi TH. Anti-

- inflammatory effects of *Achillea millefolium* on atopic dermatitis-like skin lesions in NC/Nga mice. *Am J Chin Med*. 2020; 48(05): 1121-1140. DOI: [10.1142/S0192415X2050055X](https://doi.org/10.1142/S0192415X2050055X)
119. Burk DR, Cichacz ZA, and Daskalova SM. Aqueous extract of *Achillea millefolium* L. (*Asteraceae*) inflorescences suppresses lipopolysaccharide-induced inflammatory responses in RAW 264.7 murine macrophages. *J Med Plants Res*. 2010; 4(3): 225-234. DOI: [10.1016/j.jep.2009.09.026](https://doi.org/10.1016/j.jep.2009.09.026)
  120. Raeisi H, Azimirad M, Asadi-Sanam S, Asadzadeh Aghdaei H, Yadegar A, and Zali MR. The anti-inflammatory and anti-apoptotic effects of *Achillea millefolium* L. extracts on *Clostridioides difficile* ribotype 001 in human intestinal epithelial cells. *BMC Complement Med Ther*. 2024; 24(1): 37. DOI: [10.1186/s12906-024-04335-2](https://doi.org/10.1186/s12906-024-04335-2)
  121. Fierascu I, Ungureanu C, Avramescu SM, Fierascu RC, Ortan A, Soare LC, et al. *In vitro* antioxidant and antifungal properties of *Achillea millefolium* L. *Rom Biotechnol Lett*. 2015; 20(4): 10626-10636.
  122. Georgieva L, Gadjalova A, Mihaylova D, and Pavlov A. *Achillea millefolium* L. - phytochemical profile and *in vitro* antioxidant activity. *Int Food Res J*. 2015; 22(4): 1347-1354.
  123. Kazemi M. Phytochemical and antioxidant properties of *Achillea millefolium* from the eastern region of Iran. *Int J Food Prop*. 2015; 18(10): 2187-2192. DOI: [10.1080/10942912.2014.966388](https://doi.org/10.1080/10942912.2014.966388)
  124. Arias-Durán L, Estrada-Soto S, Hernández-Morales M, Chávez-Silva F, Navarrete-Vázquez G, León-Rivera I, et al. Tracheal relaxation through calcium channel blockade of *Achillea millefolium* hexanic extract and its main bioactive compounds. *J Ethnopharmacol*. 2020; 253: 112643. DOI: [10.1016/j.jep.2020.112643](https://doi.org/10.1016/j.jep.2020.112643)
  125. Al-Ezzy RM, Al Anee R, and Ibrahim NA. Assessments of immunological activity of *Achillea millefolium* methanolic extract on albino male mice. *J Pharm Pharmacol*. 2018; 6: 563-569. DOI: [10.17265/2328-2150/2018.06.002](https://doi.org/10.17265/2328-2150/2018.06.002)
  126. Freysdottir J, Logadottir OT, Omarsdottir SS, Vikingsson A, and Hardardottir I. A polysaccharide fraction from *Achillea millefolium* increases cytokine secretion and reduces activation of Akt, ERK and NF- $\kappa$ B in THP-1 monocytes. *Carbohydr Polym*. 2016; 143: 131-138. DOI: [10.1016/j.carbpol.2016.02.017](https://doi.org/10.1016/j.carbpol.2016.02.017)
  127. Kooti W, Moradi M, Ali-Akbari S, Sharafi-Ahvazi N, Asadi-Samani M, and Ashtary-Larky D. Therapeutic and pharmacological potential of *Foeniculum vulgare* Mill: a review. *J HerbMed Pharmacol*. 2015; 4(1): 1-9.
  128. Miraj S and Kiani S. Study of antibacterial, antimycobacterial, antifungal, and antioxidant activities of *Foeniculum vulgare*: A review. *Der Pharm Lett*. 2016; 8(9): 200-205.
  129. Yang IJ, Lee DU, and Shin HM. Anti-inflammatory and antioxidant effects of coumarins isolated from *Foeniculum vulgare* in lipopolysaccharide-stimulated macrophages and 12-O-tetradecanoylphorbol-13-acetate-stimulated mice. *Immunopharmacol Immunotoxicol*. 2015; 37(3): 308-317. DOI: [10.3109/08923973.2015.1038751](https://doi.org/10.3109/08923973.2015.1038751)
  130. Lee HS, Kang P, Kim KY, and Seol GH. *Foeniculum vulgare* Mill. Protects against lipopolysaccharide-induced acute lung injury in mice through ERK-dependent NF- $\kappa$ B activation. *Korean J Physiol Pharmacol*. 2015; 19(2): 183-189. DOI: [10.4196/kjpp.2015.19.2.183](https://doi.org/10.4196/kjpp.2015.19.2.183)
  131. Salami M, Rahimalek M, and Ehtemam MH. Inhibitory effect of different fennel (*Foeniculum vulgare*) samples and their phenolic compounds on formation of advanced glycation products and comparison of antimicrobial and antioxidant activities. *Food Chem*. 2016; 213: 196-205. DOI: [10.1016/j.foodchem.2016.06.070](https://doi.org/10.1016/j.foodchem.2016.06.070)
  132. Ahmed AF, Shi M, Liu C, and Kang W. Comparative analysis of antioxidant activities of essential oils and extracts of fennel (*Foeniculum vulgare* Mill.) seeds from Egypt and China. *Food Sci Hum Wellness*. 2019; 8(1): 67-72. DOI: [10.1016/j.fshw.2019.03.004](https://doi.org/10.1016/j.fshw.2019.03.004)
  133. Boskabady M and Khatami A. Relaxant effect of *Foeniculum vulgare* on isolated guinea pig tracheal chains. *Pharm Biol*. 2003; 41(3): 211-215. DOI: [10.1076/phbi.41.3.211.15095](https://doi.org/10.1076/phbi.41.3.211.15095)
  134. Rehman NU, Ansari MN, Samad A, and Ahmad W. *In Silico* and *Ex Vivo* studies on the spasmolytic activities of fenchone using isolated guinea pig trachea. *Molecules*. 2022; 27(4): 1360. DOI: [10.3390/molecules27041360](https://doi.org/10.3390/molecules27041360)
  135. Shanmugakumar S, Gunasekaran S, Hyma P, Anil G, Praveen A, and Rajanikanth D. Phytochemical and antitubercular screening of the leaf extracts of *Foeniculum vulgare*. *World J Pharma Res*. 2013; 2(5): 1617-1625.
  136. Esquivel-Ferriño PC, Favela-Hernández JM, Garza-González E, Waksman N, Ríos MY, and Camacho-Corona MdR. Antimycobacterial activity of constituents from *Foeniculum vulgare* var. dulce grown in Mexico. *Molecules*. 2012; 17(7): 8471-8482. DOI: [10.3390/molecules17078471](https://doi.org/10.3390/molecules17078471)
  137. Yu Y, Liu G, Piao M, Lang M, Wang Y, Jin M, et al. Chemical constituents of *Polygonum aviculare* L. and their chemotaxonomic significance. *Biochem Syst Ecol*. 2022; 105: 104529. DOI: [10.1016/j.bse.2022.104529](https://doi.org/10.1016/j.bse.2022.104529)
  138. Benrahou K, Driouech M, El Guourrami O, Mrabti HN, Cherrah Y, and El Abbes Faouzi M. Medicinal uses, phytochemistry, pharmacology, and taxonomy of *Polygonum aviculare* L.: A comprehensive review. *Med Chem Res*. 2023; 32(3): 409-423. DOI: [10.1007/s00044-023-03021-1](https://doi.org/10.1007/s00044-023-03021-1)
  139. Granica S, Czerwińska ME, Żyżyńska-Granica B, and Kiss AK. Antioxidant and anti-inflammatory flavonol glucuronides from *Polygonum aviculare* L. *Fitoterapia*. 2013; 91: 180-188. DOI: [10.1016/j.fitote.2013.08.026](https://doi.org/10.1016/j.fitote.2013.08.026)
  140. Mureşan M, Olteanu D, Filip GA, Clichici S, Baldea I, Jurca T, et al. Comparative study of the pharmacological properties and biological effects of *polygonum aviculare* L. Herba extract-entrapped liposomes versus quercetin-entrapped liposomes on doxorubicin-induced toxicity on HUVECs. *Pharmaceutics*. 2021; 13(9): 1418. DOI: [10.3390/pharmaceutics13091418](https://doi.org/10.3390/pharmaceutics13091418)
  141. Mahnashi MH, Alyami BA, Alqahtani YS, Alqarni AO, Jan MS, Hussain F, et al. Antioxidant molecules isolated from edible prostrate knotweed: Rational derivatization to produce more potent molecules. *Oxid Med Cell Longev*. 2022; 2022: 3127480. DOI: [10.1155/2022/3127480](https://doi.org/10.1155/2022/3127480)
  142. Wu L, Chen Z, Li S, Wang L, and Zhang J. Eco-friendly and high-efficient extraction of natural antioxidants from *Polygonum aviculare* leaves using tailor-made deep eutectic solvents as extractants. *Sep Purif Technol*. 2021; 262: 118339. DOI: [10.1016/j.seppur.2021.118339](https://doi.org/10.1016/j.seppur.2021.118339)
  143. Mahmoudi M, Abdellaoui R, Feki E, Boughalleb F, Zaidi S, and Nasri N. Analysis of *Polygonum aviculare* and *Polygonum maritimum* for minerals by flame atomic absorption spectrometry (FAAS), polyphenolics by high-performance liquid chromatography-electrospray ionization-mass spectrometry (HPLC-ESI-MS), and antioxidant properties by spectrophotometry. *Anal Lett*. 2021; 54(18): 2940-2955. DOI: [10.1080/00032719.2021.1906267](https://doi.org/10.1080/00032719.2021.1906267)
  144. Louati K and Berenbaum F. Fatigue in chronic inflammation-a link to pain pathways. *Arthritis Res Ther*. 2015; 17: 254. DOI: [10.1186/s13075-015-0784-1](https://doi.org/10.1186/s13075-015-0784-1)
  145. Park SH, Jang S, Son E, Lee SW, Park SD, Sung Y-Y, et al. *Polygonum aviculare* L. extract reduces fatigue by inhibiting neuroinflammation in restraint-stressed mice. *Phytomedicine*. 2018; 42: 180-189. DOI: [10.1016/j.phymed.2018.03.042](https://doi.org/10.1016/j.phymed.2018.03.042)
  146. El-Far AH, Oyinloye BE, Sepehrimanesh M, Allah MAG, Abu-Reidah I, Shaheen HM, et al. Date Palm (*Phoenix dactylifera*): Novel findings and future directions for food and drug discovery. *Curr Drug Discov Technol*. 2019; 16(1): 2-10. DOI: [10.2174/1570163815666180320111937](https://doi.org/10.2174/1570163815666180320111937)
  147. Tahvilzadeh M, Hajimahmoodi M, and Rahimi R. The role of date palm (*Phoenix dactylifera* L.) Pollen in fertility: A comprehensive review of current evidence. *J Evid Based Complementary Altern Med*. 2015; 21(4): 320-324. DOI: [10.1177/2156587215609851](https://doi.org/10.1177/2156587215609851)
  148. Sassi CB, Talbi W, Ghazouani T, Amara SB, and Fattouch S. Date palm. *Nutr Compos Antioxid Prop Fruits Veg*. 2020; 681-694. DOI: [10.1016/B978-0-12-812780-3.00042-8](https://doi.org/10.1016/B978-0-12-812780-3.00042-8)
  149. Kehili HE, Zerizer S, Beladjila KA, and Kabouche Z. Anti-inflammatory effect of Algerian date fruit (*Phoenix dactylifera*). *Food Agric Immunol*. 2016; 27(6): 820-829. DOI: [10.1080/09540105.2016.1183597](https://doi.org/10.1080/09540105.2016.1183597)
  150. Hmidani A, Bourkhis B, Khouya T, Ramchoun M, Filali-Zegzouti Y, and Alem C. Phenolic profile and anti-inflammatory activity of four Moroccan date (*Phoenix dactylifera* L.) seed varieties. *Heliyon*. 2020; 6(2): e03436. DOI: [10.1016/j.heliyon.2020.e03436](https://doi.org/10.1016/j.heliyon.2020.e03436)
  151. El Hilaly J, Ennassir J, Benlyas M, Alem C, Amarouch M-Y, and Filali-Zegzouti Y. Anti-inflammatory properties and phenolic profile of six Moroccan date fruit (*Phoenix dactylifera* L.) varieties. *J King Saud Univ Sci*. 2018; 30(4): 519-526. DOI: [10.1016/j.jksus.2017.08.011](https://doi.org/10.1016/j.jksus.2017.08.011)
  152. Roshankhah S, Abdolmaleki A, and Salahshoor MR. Anti-inflammatory, anti-apoptotic, and antioxidant actions of Middle Eastern *Phoenix dactylifera* extract on mercury-induced hepatotoxicity *in vivo*. *Mol Biol Rep*. 2020; 47(8): 6053-6065. DOI: [10.1007/s11033-020-05680-4](https://doi.org/10.1007/s11033-020-05680-4)
  153. Ramchoun M, Alem C, Ghafoor K, Ennassir J, and Zegzouti YF. Functional composition and antioxidant activities of eight Moroccan date fruit varieties (*Phoenix dactylifera* L.). *J Saudi Soc Agric Sci*. 2017;

- 16(3): 257-264. DOI: [10.1016/j.jssas.2015.08.005](https://doi.org/10.1016/j.jssas.2015.08.005)
154. Alem C, Ennassir J, Benlyas M, Mbark AN, and Zegzouti YF. Phytochemical compositions and antioxidant capacity of three date (*Phoenix dactylifera* L.) seeds varieties grown in the South East Morocco. J Saudi Soc Agric Sci. 2017; 16(4): 350-357. DOI: [10.1016/j.jssas.2015.11.002](https://doi.org/10.1016/j.jssas.2015.11.002)
155. Osman NN and Al-Shubailly F. Anti-inflammatory, immune-modulatory and antioxidant effects of date fruit (*Phoenix dactylifera*) extract in rats treated with AlCl<sub>3</sub>. Int J Pharm Res Allied Sci. 2017; 6(2): 78-89.
156. Almatroodi SA, Khan AA, Aloliqi AA, Ali Syed M, and Rahmani AH. Therapeutic potential of Ajwa dates (*Phoenix dactylifera*) extract in prevention of Benzo(a)pyrene-induced lung injury through the modulation of oxidative stress, inflammation, and cell signalling molecules. Appl Sci. 2022; 12(13): 6784. DOI: [10.3390/app12136784](https://doi.org/10.3390/app12136784)
157. Bahri S, Abdennabi R, Mlika M, Neji G, Jameleddine S, and Ali RB. Effect of *Phoenix dactylifera* L. sap against bleomycin-induced pulmonary fibrosis and oxidative stress in rats: phytochemical and therapeutic assessment. Nutr Cancer. 2019; 71(5): 781-791. DOI: [10.1080/01635581.2018.1521442](https://doi.org/10.1080/01635581.2018.1521442)
158. Ljubuncic P, Dakwar S, Portnaya I, Cogan U, Azaizeh H, and Bomzon A. Aqueous extracts of *Teucrium polium* possess remarkable antioxidant activity *in vitro*. Evid Based Complement Alternat Med. 2006; 3(3): 329-338. DOI: [10.1093/ecam/nel028](https://doi.org/10.1093/ecam/nel028)
159. Bahramikia S and Yazdanparast R. Phytochemistry and medicinal properties of *Teucrium polium* L. (Lamiaceae). Phytother Res. 2012; 26(11): 1581-1593. DOI: [10.1002/ptr.4617](https://doi.org/10.1002/ptr.4617)
160. Khazaei M, Nematollahi-Mahani SN, Mokhtari T, and Sheikhbahaei F. Review on *Teucrium polium* biological activities and medical characteristics against different pathologic situations. J Contemp Med Sci. 2018; 4(1): 1-7. DOI: [10.22317/jcms.03201801](https://doi.org/10.22317/jcms.03201801)
161. Amraei M, Ghorbani A, Seifinejad Y, Mousavi SF, Mohamadpour M, and Shirzadpour E. The effect of hydroalcoholic extract of *Teucrium polium* L. on the inflammatory markers and lipid profile in hypercholesterolemic rats. J Inflamm Res. 2018; 11: 265-272. DOI: [10.2147/JIR.S165172](https://doi.org/10.2147/JIR.S165172)
162. Rahmouni F, Hamdaoui L, and Rebai T. *In vivo* anti-inflammatory activity of aqueous extract of *Teucrium polium* against carrageenan-induced inflammation in experimental models. Arch Physiol Biochem. 2017; 123(5): 313-321. DOI: [10.1080/13813455.2017.1333517](https://doi.org/10.1080/13813455.2017.1333517)
163. Al-Naemi HA, Alasmar RM, and Al-Ghanim K. Alcoholic extracts of *Teucrium polium* exhibit remarkable anti-inflammatory activity: *In vivo* study. Biomol Biomed. 2024; 24(1): 82-92. DOI: [10.17305/bb.2023.9239](https://doi.org/10.17305/bb.2023.9239)
164. Ait Chaouche FS, Mouhouche F, and Hazzit M. Antioxidant capacity and total phenol and flavonoid contents of *Teucrium polium* L. grown in Algeria. Mediterr J Nutr Metab. 2018; 11(2): 135-144. DOI: [10.3233/MNM-17189](https://doi.org/10.3233/MNM-17189)
165. Özer Z, Kılıç T, Çarıkçı S, and Yılmaz H. Investigation of phenolic compounds and antioxidant activity of *Teucrium polium* L. decoction and infusion. Balıkesir Üniv Fen Bilim Enst Derg. 2018; 20(1): 212-218. DOI: [10.25092/baunfbed.370594](https://doi.org/10.25092/baunfbed.370594)
166. El Atki Y, Aouam I, Taroq A, Lyoussi B, Taleb M, and Abdellaoui A. Total phenolic and flavonoid contents and antioxidant activities of extracts from *Teucrium polium* growing wild in Morocco. Mater Today Proc. 2019; 13: 777-783. DOI: [10.1016/j.matpr.2019.04.040](https://doi.org/10.1016/j.matpr.2019.04.040)
167. Matic S, Popovic S, Baskic D, Todorovic D, Vukovic N, Stankovic M, et al. Methanolic extract of *Teucrium Polium* exerts immunomodulatory properties in human peripheral blood mononuclear cells. Exp Appl Biomed Res. 2022; 23(4): 345-351. DOI: [10.2478/sjecr-2020-0018](https://doi.org/10.2478/sjecr-2020-0018)