












Systematic Review

Exploiting Anti-tumor Effects of *Salmonella typhimurium*: A Systematic Review

Seyed Alireza Taheri¹ , Mahsa Norouzi² , Atefehsadat Monirvaghefi³ , Fatemeh Najafi⁴ , Abdolmahdi Asfaram Meshkinshahr⁵ , Sara Aghili⁶ , Golnaz Behzad⁷ , Dorsa Mousavi Khatibi⁸ , Bahare Kasaei⁹ , and Armin Batmani^{10,*} 

¹ Faculty of Pharmacy, Mashhad University of Medical Sciences, Mashhad, Iran

² Faculty of Pharmacy, Tabriz University of Medical Sciences, Tabriz, Iran

³ Center for Drug Delivery and Nanomedicine, Department of Pharmaceutical Sciences, University of Nebraska Medical Center, Omaha, Nebraska, United States

⁴ Faculty of Pharmacy, Tehran Medical Sciences, Islamic Azad University, Tehran, Iran

⁵ Pharm-D, Faculty of Pharmacy, Azerbaijan Medical University, Azerbaijan

⁶ Pharm-D, Faculty of Pharmacy, Rajiv Gandhi University of Health Sciences, Karnataka, India

⁷ Faculty of Pharmacy, Lorestan University of Medical Sciences, Khorramabad, Iran

⁸ Faculty of Pharmacy, Ardabil University of Medical Science, Ardabil, Iran

⁹ Faculty of Pharmacy, Tehran Medical Sciences, Islamic Azad University, Tehran, Iran

¹⁰ Pharm-D, Faculty of Pharmacy, Yaroslav the Wise Novgorod State University, Novgorod, Russia

* **Corresponding author:** Armin Batmani, Faculty of Pharmacy, Yaroslav the Wise Novgorod State University, Novgorod, Russia. Email: armin.batmaniasl@gmail.com

ARTICLE INFO

Article History:

Received: 10/09/2023

Revised: 01/10/2023

Accepted: 05/10/2023

Published: 25/10/2023



Keywords:

Anti-tumor properties

Cancer therapy

In vivo

Salmonella typhimurium

Tumor targeting

ABSTRACT

Introduction: *Salmonella typhimurium* (*S. typhimurium*) has emerged as a promising agent for cancer therapy. This systematic review aimed to comprehensively analyze the existing literature regarding the utilization of *S. typhimurium* as a therapeutic strategy against cancer. The present systematic review aimed to evaluate the current state of knowledge regarding the anti-tumor properties of *S. typhimurium*, encompassing its tumor-targeting mechanisms, impact on tumor growth, modulation of the tumor microenvironment, and potential for combination therapies.

Materials and methods: A systematic literature search was conducted across major scientific databases, including PubMed, Web of Science, and Scopus, using predefined search terms. Studies published between 2000 and 2023 were included if they investigated the anti-tumor effects of *S. typhimurium in vivo*. Studies were independently screened, selected, and evaluated for quality by two experts in the field.

Results: The systematic review identified 152 relevant studies that met the inclusion criteria. These studies collectively demonstrated the ability of *S. typhimurium* to selectively target and colonize tumors, resulting in significant tumor growth inhibition in various cancer types. Mechanistic insights revealed that *S. typhimurium* can induce direct cytotoxicity, modulate the tumor microenvironment, and activate anti-tumor immune responses. Additionally, studies highlighted the potential of combining *S. typhimurium* with conventional therapies or immune checkpoint inhibitors to enhance therapeutic efficacy.

Conclusion: This systematic review underscores the promising potential of *S. typhimurium* as a novel and multifaceted approach to cancer therapy. The accumulated evidence suggests that *S. Typhimurium* possesses inherent tumor-targeting capabilities, exerts direct anti-tumor effects, and can synergize with other treatment modalities.

1. Introduction

One of the most challenging health problems to treat is cancer, which is a leading cause of death worldwide¹. On

average, 10 million people die from cancer each year. Detecting and treating cancer at an early stage is crucial. It

► **Cite this paper as:** Taheri SA, Norouzi M, Monirvaghefi A, Najafi F, Asfaram Meshkinshahr A, Aghili S, Behzad G, Mousavi Khatibi D, Kasaei B, Batmani A. Exploiting Anti-tumor Effects of *Salmonella typhimurium*: A Systematic Review. Journal of Lab Animal Research. 2023; 2(5): 51-62. DOI: 10.58803/jlar.v2i5.30



The Author(s). Published by Rovedar. This is an open-access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

is important to note that most conventional treatments can damage healthy tissues, even though they are lifesaving². For example, chemotherapeutics, surgeons, and radiologists effectively save lives³. Surgical removal can be used to treat certain cancer types and stages. In addition, metastases and relapse are possible with this method⁴. The success of chemotherapy and radiotherapy in treating cancer varies accordingly, especially distant recurrences of tumors and adverse effects. The development of new cancer treatment ideas and strategies is crucial for improving the effectiveness in patients⁵⁻⁷. According to several studies, tumor treatment failure can be attributed to such regions^{8,9}. Furthermore, tumor-specific therapeutic agents cannot be delivered to tumors because of abnormal vascular architecture. Various organisms, including fungi, and pathogens (parasitic worms [hydatid cyst protoscolex, *Trichinella spiralis*] or even protozoan (*Trypanosoma cruzi*) offer potential anti-cancer properties through bioactive compounds and immunomodulation¹⁰⁻¹⁴. *Escherichia coli*, *Clostridium*, *Salmonella*, and *Bifidobacterium*, among others, contain inherent characteristics that make them tumor-targeting and tumor-killing bacteria¹⁵.

It has long been recognized that *Salmonella typhimurium* (*S. typhimurium*) is a significant cause of foodborne illness in humans, which is a gram-negative bacterium from the *Enterobacteriaceae* family^{16,17}. This pathogenic bacterium has recently been identified as a potential cancer therapeutic agent¹⁸. Investigations indicate *S. typhimurium* enters and selectively kills solid tumor tissues^{19,20}. However, healthy cells were not harmed; a novel and potent tool for treating cancer has been discovered in *S. typhimurium* based on the intriguing discovery²¹. The ability of some bacteria to home tumors has been demonstrated while heterologous genes have been delivered intracellularly by other invasive species^{22,23}. It is more advantageous to express target genes in *S. typhimurium* since they are highly replicating and invasive²⁴. A tumor hypoxic zone promotes the survival of optional anaerobes that act as anti-tumor agents²⁵. The therapeutic gene must be delivered effectively to the target tissues or cells for gene therapy to be successful²⁶. *S. typhimurium* is one of the greatest pathogens for bacteria-mediated cancer treatments (BMCT). Animals with highly aerobic conditions can spread it systemically, and a hypoxic tumor region is its preferred colonization site, where it eventually settles. In addition to conventional therapies, *S. Typhimurium* can colonize hypoxic and necrotic and metastatic tumors^{27,28}.

The objective of this systematic review is to evaluate the current state of knowledge regarding the antitumor properties of *S. typhimurium*, encompassing its tumor-targeting mechanisms, impact on tumor growth, modulation of the tumor microenvironment, and potential for combination therapies in animal models. *Salmonella*'s potential as a key microbial agent in cancer therapy will be investigated, as well as engineering methods that can be used to create *Salmonella*-based cancer treatments of the future.

2. Materials and Methods

The article presents a review of studies published

between the years 2000 and 2022 in order to come up with a conclusion. All relevant studies on *S. typhimurium*'s anti-cancer properties are identified through systematic and exhaustive literature searches. A comprehensive search of multiple electronic databases, such as PubMed, Scopus, Web of Science, and Database, will begin the review process. The search is supplemented with manual searches of reference lists and relevant journals to minimize the risk of missing relevant studies. Searching keywords were *Salmonella typhimurium*, *Salmonella*-based therapy, *Salmonella*, cancer therapy, cancer, immunotherapy, gene therapy, and *in vivo*.

2.1. Inclusion and exclusion criteria

Upon retrieving the studies, screening was conducted in two stages. Reviewing the titles and abstracts of the papers was the first step in the screening process. Second, the quality of the articles was evaluated based on the full text of their studies. The title and abstract screening procedures involved studies with predefined inclusion, and the exclusion criteria led to the removal of irrelevant or duplicates. The full texts of studies were reviewed once they were available.

2.2. Quality assessment and data extraction

This study was developed based on a standardized data extraction form to extract relevant information from the selected studies. It included publication year, authors, sample size, and experimental model, which are the most important characteristics of the study. *Salmonella typhimurium* has a wide range of anti-tumor effects, including apoptosis induction, immune response modulation, tumor targeting, tumor growth inhibition, and other mechanisms. Several appropriate tools were used to assess the inclusion of studies, including the Cochrane Collaboration's Risk of Bias tool for randomized controlled trials for non-randomized studies. Upon the completion of independent quality assessments by both experts, the inconsistencies between the two were discussed.

2.3. Data analysis

Data synthesis and analysis were based on a narrative framework since the included studies are heterogeneous. A descriptive summary of the evidence aimed to provide a comprehensive overview. The results were arranged and presented according to the type of cancers investigated.

3. Results

Anti-tumor properties of *S. typhimurium* against cancer, and a systematic review of these studies were conducted. The review included 480 studies, which met the inclusion criteria, from 530 publications. Overall, 50 of them were duplicates and were excluded. Among these, 137 full-text articles were reviewed for eligibility. As a result of removing duplicates and screening the articles for eligibility, 24 articles were included in the quantitative synthesis (Figure 1).

3.1 Anti-tumor mechanism of *Salmonella typhimurium*

3.1.1. Hypoxic environment and tumor vasculature

A tumor's blood supply is unevenly distributed and chaotic, resulting in chronic and acute hypoxia²⁹. As a result, oxygen delivery to the cells is diminished, and the proliferation of the cells is disrupted. Bacteria, such as *S. typhimurium*, thrive in hypoxic environments³⁰. An abnormal vasculature can result from tumor angiogenesis that, in comparison with normal tissue, is more vascularized³¹. It is likely that *S. typhimurium* can invade such an unruly and highly vascularizing tumor microenvironment³². These facultative anaerobic bacteria are more likely to be localized. It begins to kill tumor cells so that they can survive and take up nutrients. In addition, chaotic vasculature contributes to the disorder³³. A double-auxotrophic mutant *S. typhimurium* strain A1-R with tumor-targeting properties has been demonstrated to increase vascularity and destroy tumor blood vessels³⁴.

3.1.2. Abundant nutrients and competitive nature of *Salmonella typhimurium*

There are a variety of micronutrients in the tumor microenvironment³⁵. In a study of lung and pancreatic adenocarcinomas in mice, nutrients positively influencing

cancer cell metabolism differed between tumors and the circulation³⁶. Cancers generate energy and biomass through metabolic adaptation, which accumulates metabolites. Moreover, it alters the expression of normal genes³⁷. The tumor's microenvironment is immune-suppressive. As a result, by providing nutrients and protecting the bacteria from host immunosurveillance, these compounds could enable attenuated auxotrophic bacteria to survive and grow³⁸. It has been found that prostate cancer and breast cancer can be treated in mice models by employing auxotrophic *Salmonella* for tryptophan, arginine, and leucine³⁹.

3.1.3. Tumor penetration

Salmonella typhimurium has been chosen for BMCT due to its potential for deep tumor penetration⁴⁰. Passive transport is traditionally used to distribute chemotherapeutic drugs. *Salmonella typhimurium* invades deep tumor tissues by utilizing the abundant nutrients surrounding it. When *S. Typhimurium* has been administered systemically, apoptosis is induced by systemic subcutaneous administration⁴¹. In tumor tissue, bacterial motility influences the spatial distribution of bacteria; the motility of bacteria increases their penetration depth⁴². It has also been suggested that tumor migration of *S. typhimurium* is not dependent on motility or chemotaxis but a passive process. In order to study the

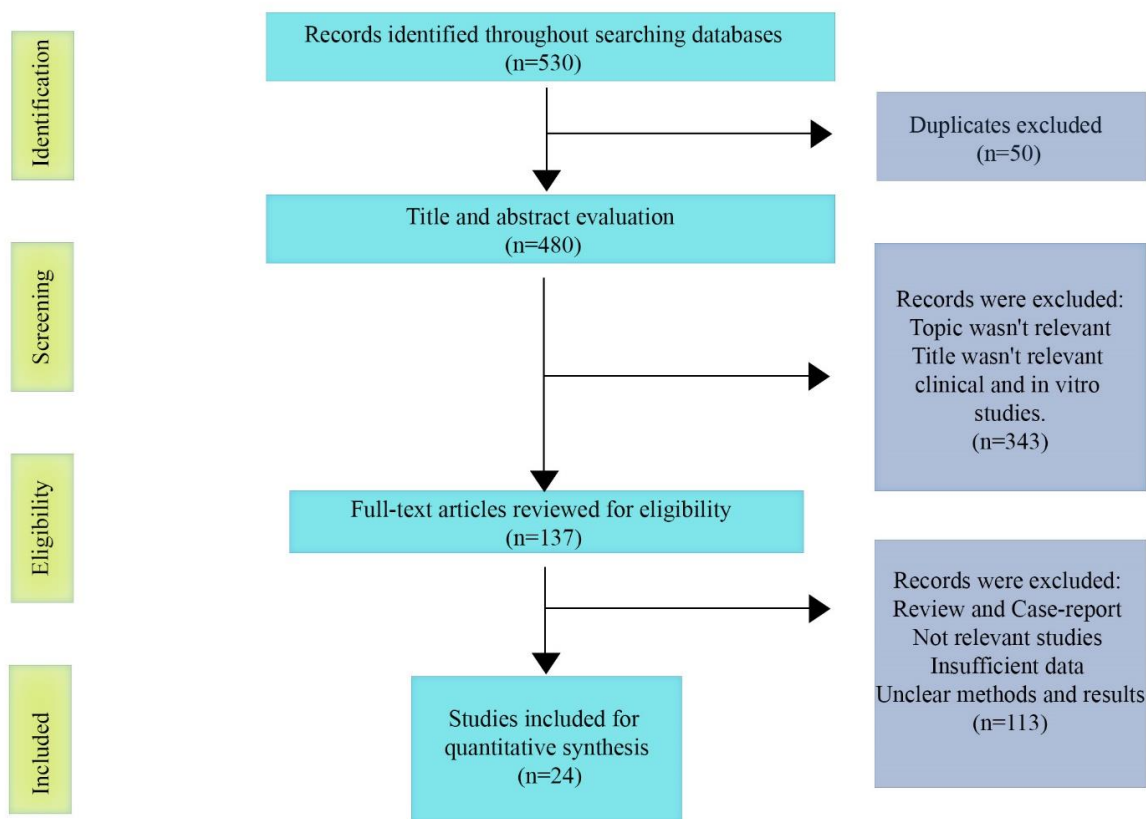


Figure 1. Methodology of the current study

events of tumor-colonization, different strains, and time points post-infection were used *in vivo*⁴³. Tumor tissue is protected from bacteria by neutrophils; bacteria colonize intratumor more readily when their numbers are diminished⁴⁴.

3.1.4. Apoptosis and autophagy-inducing intrinsic anti-tumor action

Salmonella typhimurium has been shown to directly kill cancer cells in several *in vivo* studies^{45,46}. Genetically engineered *S. typhimurium* strain A1-R infected cancer cells have grown into large colonies, as revealed by high-resolution multiphoton tomography images⁴⁷. *Salmonella typhimurium* can induce both apoptosis and autophagy when it enters the cells of cancer cells^{45,48}. There is no clear understanding of how *S. typhimurium* induces apoptosis. Bacterial toxins and competition for nutrients with cancer cells may cause apoptosis. In addition, autophagy may be induced; in tumor cells, scavengers are less active than in normal cells, meaning they are less active⁴⁹. Phospho-Protein Kinase B (P-AKT) is downregulated in an AKT-dependent manner/ The Phospho-mammalian Targets Of Rapamycin (P-mTOR) pathway regulates cellular proliferation and survival⁵⁰. Cellular physiology and homeostasis are influenced by the P-AKT / P-mTOR pathway. Matrix MetalloProteinase 9 (MMP-9) expression is reduced by the downregulation of this pathway, which is involved in metastasis; this oncoprotein plays an important role⁵¹.

3.1.5. Inhibition of angiogenesis

It is essential to understand that angiogenesis plays a crucial role in tumor progression and development, HIF-1 α and VEGF play a vital role in tumor angiogenesis⁵². The invasion of *S. typhimurium* in a tumor decreases the expression of VEGF and HIF-1 α , inhibiting tumor angiogenesis by activating the P-AKT/P-mTOR pathway⁵³. A recently identified tumor angiogenesis inhibitor protein is another mechanism for suppressing angiogenesis; through HIF-1, Connexin 43 (Cx43) inhibits VEGF expression besides interfering with its composition⁵⁴. A tumor-targeting double-auxotrophic mutant *S. typhimurium* strain A1-R shows an association between a higher tumor vasculature and the destruction of tumor blood vessels⁵⁵.

3.1.6. Immunomodulation in tumor tissue

In animal models, *S. typhimurium* can manipulate the immune components of the tumor function to inhibit tumor growth by changing the tumor microenvironment from an immunosuppressive to an immunogenic state⁵⁶. Evidence shows that *S. typhimurium* infection increases macrophage, natural killer (NK), CD4+ helper T cells, and CD8+ cytotoxic T cell infiltration⁵⁷. Colony-Stimulating Factor 1 (CSF-1) and Chemokine C-C motif chemokine Ligand 2 (CCL-2) are chemo-attractants released by tumor cells in order to recruit monocytes that undergo differentiation into

macrophages of the M2 subtype^{58,59}. Tumor growth and malignancy are promoted by M2 macrophage polarization, which secretes immune-suppressive molecules to suppress antitumor immune responses in the host⁶⁰. As a result of *S. typhimurium* invasion, these cytokines are released, including IL-10 and arginase 1 (Arg1)^{61,62}. It has been demonstrated that TAMs activate M1 macrophages by releasing various activation markers, such as Sca-1 and MHC class II, in response to *S. typhimurium* invasion⁶³. This is a paradigm shift from M2 to M1. The M1 macrophages orchestrate the anti-tumor immune responses by expressing nitric oxide synthase (NOS2) and TNF- α , which enhance the protective immune response to tumors *in vivo*⁴¹.

A T cell that inhibits cytotoxic T-lymphocytes specific for tumor antigens mitigates antitumor immunity⁶⁴. When injected into a colon cancer model, attenuated *S. typhimurium* reduces regulatory T (Treg) cell⁶⁵. Cell surface molecule CD44, present on both cancer cells and Treg, is downregulated by *Salmonella* treatment⁶⁶. Gap junction formation can be facilitated by *S. typhimurium* infection by upregulating CX43⁶⁷. Tumor cells can present antigenic peptides to dendritic cells via gap junctions. Through antigenic presentation, cytolytic T cells can be activated against the tumor antigen, ultimately responsible for halting distant uninfected tumor growth *in vivo*⁶⁸.

3.1.7. Orchestration of tumor-associated macrophage function and polarization

The tumor-associated macrophage (TAM) is a constituent of the leukocytic infiltrate and has been used as a paradigm for cancer-related inflammation, tumor growth, invasion, metastasis, and drug resistance^{69,70}. It is well-known that M1 macrophages have anti-tumor properties, whereas M2 macrophages promote tumor development, angiogenesis, and progression⁷¹. There are two sides to TAM, exhibiting pro- and anti-tumor activity, and this cell has high immunological reprogramming potential, especially in the presence of IFN- α or IFN- γ ⁷². In a different approach, suppression of TAMs can be achieved by designing cancer vaccines against proteins which are overexpressed by TAMs, such as Legumain⁷³. Legumain encodes an asparaginyl endopeptidase that is highly upregulated in murine and human tumor tissues⁷⁴.

3.1.8. Release of cytotoxic chemicals

During *S. typhimurium* treatment, cytotoxic compounds such as granzyme and perforin are released to kill tumor cells⁷⁵. Immunomodulatory molecules, such as chemokines and cytokines, stimulate the immune system to eliminate tumors. Consequently, several cytotoxic agents, including Fas ligand (FASL), IL-18, TRAIL, IL-2, TNF- α , and cytolysin A, have been expressed in *S. typhimurium in vivo*⁷⁶⁻⁸⁰. Breast and colon tumors were inhibited by *S. typhimurium* engineered to express FASL, a proapoptotic cytokine⁸¹⁻⁸³. Moreover, murine mammary tumors expressing the cytotoxic protein HlyE under hypoxic conditions showed increased necrosis and reduced growth when said with a

hypoxia-inducible promoter⁸⁴. The RecA promoter is used in *S. typhimurium* to control the secretion of murine TRAIL, growth of mammary tumors delayed, activated apoptosis^{85,86}. Similar results were achieved by hypoxia-induced nirB-regulated TRAIL for the suppression of melanoma *in vivo*⁸⁷.

3.1.9. Role of the type III secretion system

The virulence factor of *S. typhimurium* is well known, and it is encoded by *Salmonella* pathogenicity island 1 (SPI1) as the type III secretion system (T3SS)⁸⁸. As a result, it is possible to inject effector proteins directly into host cells. Signaling pathways within cells are interfered with the cytoskeleton network in the host cell can be manipulated, and colonization is enabled⁷⁶. They can produce and store most of the effector proteins secreted by the T3SS⁸⁹. Injection of *Salmonella* pathogenicity island 2 (SPI2) effector molecules promotes cellular invasion promoted by SPI1⁹⁰. *Salmonella* is resistant to most host defense mechanisms due to their ability to reconstruct endosomes into *Salmonella*-containing vacuoles to avoid host defense mechanisms and to enable intracellular survival^{91,92}. Thus, SPI1 and SPI2 work together to facilitate the invasion of native forms or bacterial vectors⁹³. It is, therefore, possible for *S. typhimurium* to induce apoptosis in established infection, and this has been confirmed by *S. typhimurium* effector expression causing apoptosis in tumor cells through activation of caspases 3 and 7, SpvB (an ADP-ribosyl transferase enzyme)⁹⁴.

P-glycoprotein is expressed in many cancers and is downregulated by *S. typhimurium* effector SipA such as breast, kidney, colon, and lymphoma⁹⁵⁻⁹⁷. Proteins of the T3SS family, such as inner rod proteins, needles, and flagellins inner rod, are perceived by nucleotide-binding oligomerization domain (NOD)-like receptor (NLR) family apoptosis inhibitory protein (NAIP) to activate the NAIP-NLR family caspase-associated recruitment domain-containing protein 4 (NLRC4)⁹⁸. Caspases-1 and -8 are activated due to the assembly of the NLRC4 inflammasome⁹⁹. As caspase-1 is activated, pyroptosis is induced, and caspase-8 stimulates caspase-3/7, resulting in the inflammatory death of the host cells, known as PANoptosis¹⁰⁰⁻¹⁰². When effector proteins are internalized, the host immune response is modulated, cytokines are secreted, the cytoskeleton of the host cells is rearranged, and the invasion of bacteria is facilitated^{76,103}. To develop cancer vaccines, we must improve antigen delivery; a promising strategy is *Salmonella* T3SS. Mice with tumors expressing NY-ESO-1 regressed when NY-ESO-1 tumor antigen was administered through T3SS¹⁰⁴. Heterologous antigens like *Listeria monocytogenes*' MHC class I-peptide p60 were employed to translocate into host cells using T3SS from *Salmonella*, CD8 T cells induced by antigens, regression of tumors is significant¹⁰⁵. Survivin, an antigen that is transported into antigen-presenting cells by SPI2, is an effector of immune responses induced by the SseF protein of SPI2¹⁰⁶. In a mouse model, TAA was delivered via the SPI-2-regulated T3SS to stimulate anti-tumor activity (Figure 2)¹⁰⁷.

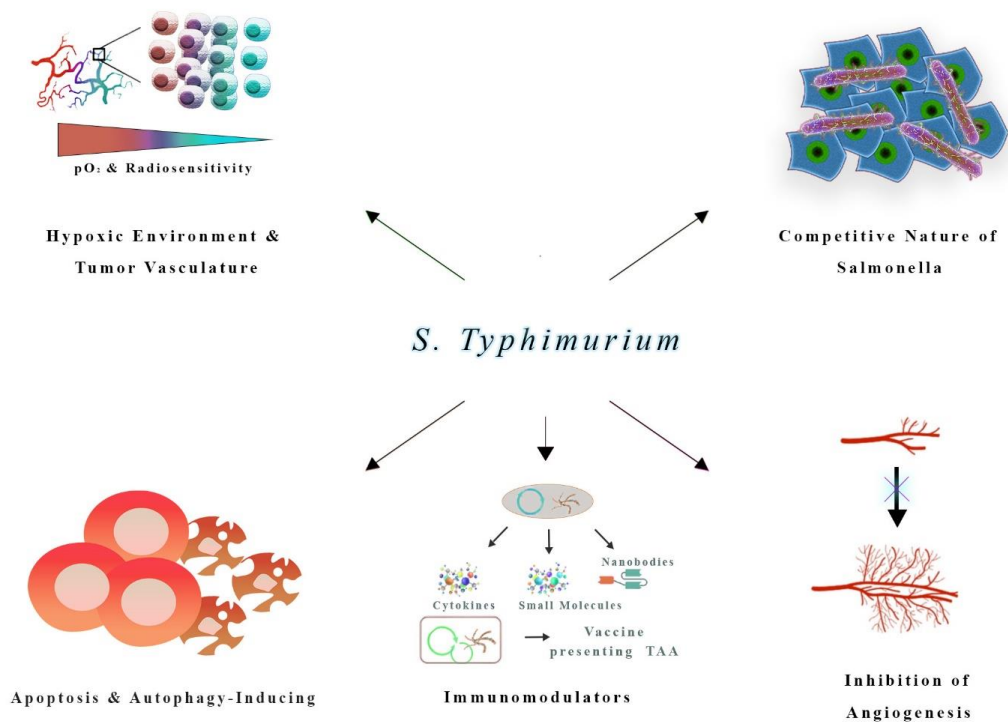


Figure 2. Anti-tumor mechanisms of *S. typhimurium*. Key strategies employed by *Salmonella typhimurium* for combating tumors include its affinity for hypoxic tumor environments and chaotic vasculature, its competitive nature in utilizing abundant nutrients, its ability to induce apoptosis and autophagy within cancer cells, its inhibition of angiogenesis, and its role in immunomodulation in the tumor microenvironment. Together, these mechanisms illustrate how *S. typhimurium* exerts a multifaceted anti-tumor action, potentially impacting tumor vasculature, nutrient availability, and immune responses to facilitate tumor reduction.

4. *Salmonella typhimurium*'s selective tumor targeting

Cancer researchers have been intrigued by *S. typhimurium*'s ability to specifically target tumor tissues¹⁰⁸. Several intricate factors will be discussed in this subsection that *S. typhimurium* has the potential to benefit cancer patients over traditional cancer therapies because of its preferential tumor homing.

Various factors contribute to the unique conditions of the tumor microenvironment¹⁰⁹. Healthy tissues do not have those characteristics. An inefficient or irregular blood supply causes low oxygen levels and vasculature leaks in tumors, providing an escape route for molecules and cells. *Salmonella* uses these proteins to selectively target tumors in both cases, leaky vasculature and hypoxia¹¹⁰.

Chemotaxis is one of the most remarkable abilities of *Salmonella*; chemical gradients can be sensed and responded to by the body¹¹¹. When it comes to targeting tumors, *Salmonella* can detect various chemotactic signals in the tumor microenvironment; bacteria are guided toward the tumor by these molecules¹¹². Several signals are generated by necrotic and hypoxia areas inside the tumor, including metabolites, specific molecules, and gradients of nutrients¹¹³.

A tumor is infected with *Salmonella* once reached; it can increase selectively within the tumor because of its nutrient-rich environment and relative protection¹¹⁴. During tumor microenvironment invasion, bacteria can take advantage of the weakened immune system to exploit available nutrients; the tumor can survive and reproduce more easily¹¹⁵.

During tumor growth, the leaky vessels allow *Salmonella* to enter the tumor tissues and extravasate from the bloodstream¹¹⁶. *Salmonella* can access the tumor due to this vascular permeability, how it can be improved, produce anti-tumor effects, and replicate them¹¹⁷. The molecular interactions between *Salmonella* and tumor cells have been demonstrated in studies. Virulence factors are expressed by certain *Salmonella* strains; it allows them to attach to receptors on tumor cells that are expressed in a preferential manner, enabling cancer cells to be targeted and internalized¹¹⁸.

Immune responses are triggered by *Salmonella*'s presence in the tumor microenvironment¹¹⁹. Immune cells can be activated by bacteria, like dendritic cells and macrophages, in the immune response against tumors; they play a critical role¹²⁰. As a result of this immunomodulatory effect, the immune system may be able to respond more aggressively to Cancer and enhance anti-tumor activity.

5. Preclinical studies and animal models

Research involving *Salmonella*-based anti-tumor therapies has shown great potential for establishing *Salmonella*'s therapeutic potential for a variety of cancers¹²¹. Animal models were used in these studies, which provide invaluable insights into how *Salmonella*

affects cancer progression and growth biologically and therapeutically. *Salmonella* has been shown to be effective in treating different kinds of cancer and in combination with existing therapies in some notable preclinical studies^{24,122}.

An animal model of breast cancer was used in recent studies¹²³. Cancer cells were specifically targeted by *Salmonella* strains that carried cytotoxic genes¹²⁴. As a result of the *S. typhimurium* being administered, there was a significant reduction in tumor size and an important tumor regression was observed in the mice treated with *Salmonella* administered intravenously, which led to prolonged survival^{125,126}. Through this method, *S. typhimurium* was demonstrated to be capable of selectively targeting and inhibiting the growth of breast cancer cells *in vivo*¹²⁷.

Another preclinical study using *Salmonella* as a vehicle for gene therapy in mice with prostate cancer was conducted¹²⁸. *Salmonella* engineered to infiltrate prostate tumors successfully activated immune responses that targeted prostate tumors^{129,130}. A *Salmonella*-based therapy could benefit prostate cancer patients by boosting anti-tumor immune responses through enhanced reduced tumor growth and tumor cell killing *in vivo*¹³¹.

A *Salmonella*-based therapy has been shown to reduce tumor burden in mouse models of colorectal cancer¹³². Anti-cancer agents expressed by *Salmonella*, such as tumor necrosis factor (TNF), to treat colorectal cancer, were used. As a result of the treatment, growth, and tumor size were significantly reduced.

In an *in vivo* study, ovarian cancer is modeled in mice, along with standard chemotherapy drugs, *Salmonella* was administered¹³³. Chemotherapy alone had no anti-tumor effects, but combination treatment had enhanced outcomes. There was a significant reduction in tumor growth and increased survival rates with the combination treatment compared to chemotherapy alone. As a result of *Salmonella*'s synergistic action with chemotherapy, combination therapies can strengthen tumor regression¹³⁴. It is important to note that immune checkpoint inhibitors have been studied in combination with *Salmonella*-based immunotherapies, which cancer-fighting drugs increase the immune system's ability to fight cancer¹³⁵. *Salmonella*-based treatment and immune checkpoint inhibitors have been shown to significantly reduce tumor size and prolong survival in preclinical studies in mouse models of melanoma^{136,137}. Immunotherapies can be enhanced and complemented by *Salmonella* in this way¹³⁸.

6. Challenges

Live-engineered bacteria represent a unique therapeutic opportunity that is accompanied by a number of challenges¹³⁹. First, genes or motile genetic elements confer antibiotic resistance to living naturally modified bacteria. It is safer and more stable to engineer the expression cassette using chromosomal integration

without antibiotic selection markers¹⁴⁰. Second, the manufacturing of GMP-grade test articles presents a specific challenge because live bacteria can neither be sterilized by heating nor filtered¹⁴¹. Furthermore, there would be no way to test for sterility by the conventional regulatory standard. In order to ensure "sterility," dedicated clean rooms with strict aseptic protocols and frequent monitoring of the process must be used during purification and production¹⁴². However, the final products need to be tested to ensure that no other diseases or conditions are present, even though they cannot be proven to be sterile. The third reason is that live bacteria proliferate in target tissues and thus spread¹⁴³. It is not necessary that the administered dose corresponds to the effective dose (toxic or therapeutic). Target tissue "quality" influences the effective dose more than anything else which is determined by accessibility, whether there is hypoxia/tumor necrosis, and if pre-existing inflammatory cells have infiltrated the tumor¹⁴⁴. Bacteria, at low doses, particularly when administered systemically, can harm the immune system¹⁴⁵. It is less predictable and can take much longer for an infection to establish itself in the target tissue. Over time, the patients may become less vigilant, posing a greater risk. Fourth, oncolytic therapy involves turning a tumor into an infection that destroys the cancer; this could lead to severe consequences without proper management. It is essential to strike a carefully calculated balance between an infection's therapeutic benefits and toxic side effects. The practical difficulty of achieving this is great, antibiotics are effective only when administered on time, which would allow the infection to be eradicated before the anti-tumor effect was achieved¹⁴⁶. A systemic inflammatory response risk during a late intervention is much higher. A multidisciplinary approach is needed to manage therapeutic infections, including oncologists, Surgeons, interventional radiologists, or infectious disease specialists need to manage conditions requiring invasive treatment¹⁴². If an intratumoral infection is established, it is the team's responsibility to decide what to do and when to intervene. Fifth, in clinical settings, live biological agents are used. Public health and the environment are always concerns when it comes to its potential impacts¹⁴⁷.

7. Future and prospect

Researchers are working to better understand *Salmonella*'s mechanisms and address its weaknesses in order to develop future anti-tumor therapies based on *Salmonella*¹²³. As a result of personalized medicine approaches, *salmonella*-based anti-tumor treatments may become a major part of the future treatment of cancer¹⁴¹. Personalized medicine incorporates genetics, molecular biology, and immunity into treatment planning. *Salmonella*'s genetic engineering capability allows precise therapeutic strategies. *Salmonella*, which can be genetically engineered to produce antigens and receptors specific to cancer cells, can be used to selectively kill cancer cells without harming healthy cells¹³⁹.

Research can optimize *Salmonella*'s anti-tumor properties by changing its genetic makeup; by enhancing its apoptotic capability, researchers could improve its immune-stimulating properties for different cancer types¹¹⁷. The presence of specific biomarkers can indicate the efficacy of *Salmonella*-based therapies. This approach selects patients based on their biomarkers to optimize treatment efficacy and minimize side effects^{48,57}.

When *Salmonella*-based treatments are combined with other immunotherapies, they can be more effective at treating cancer; they may produce a synergistic effect. This combination approach boosts the immune system and overcomes immunosuppressive microenvironments to prevent tumors from spreading. *Salmonella*-based therapies can combine immune checkpoint inhibitors with immune checkpoint inhibitors when the immune system encounters cancer cells, it attacks them¹²². *Salmonella* can enhance antitumor responses by activating immune responses and modulating the tumor microenvironment.

CAR-T cell therapy targets specific cancer antigens, with T cells engineered to target those antigens. It is more likely that CAR-T cells will enhance anti-cancer activity and tumor infiltration when delivered directly to tumors using *Salmonella*. A cancer vaccine is a personalized treatment because it incorporates tumor antigens into *Salmonella*⁶⁵. Vaccines may help identify and destroy cancer cells by stimulating the immune system¹⁰⁴.

8. Conclusion

The current study has unveiled a groundbreaking frontier in the battle against cancer by harnessing the remarkable antitumor properties of *S. typhimurium*. Through a series of innovative *in vivo* studies, the current study has demonstrated the unprecedented potential of this bacterium as a novel therapeutic agent. This study not only shed light on the underlying mechanisms of *Salmonella*'s anti-cancer effects but also paved the way for the development of cutting-edge treatments that exploit this unique biological weapon. This exciting discovery holds promise for a future where *S. typhimurium* stands as a formidable ally in the fight against cancer, offering new hope and novel strategies for improving patient outcomes.

Declarations

Competing interests

The authors declare no conflict of interest.

Authors' contributions

The conceptualization of this project was spearheaded by Armin Batmani, who played a pivotal role in its development. The task of writing and preparing the original draft involved the dedicated efforts of Seyed Alireza Taheri, Mahsa Norouzi, Atefehsadat Monirvaghefi, Fatemeh Najafi, Abdolmahdi Asfaram Meshkinshahr, and

Sara Aghili. The project underwent a rigorous review and editing process, with contributions from Golnaz Behzad, Dorsa Mousavi Khatibi, and Bahare Kasaei. The project was self-funded, and it was executed under the supervision and guidance of Armin Batmani. All authors checked and approved the final version of the manuscript for publication in the present journal.

Funding

No fund.

Ethical considerations

All applicable international, national, and/or institutional guidelines for the care and use of animals were followed.

Availability of data and materials

The datasets generated during and/or analyzed during the current study are available from the corresponding author upon reasonable request.

Acknowledgments

None.

References

1. Aganja RP, Sivasankar C, Senevirathne A, and Lee JH. Salmonella as a promising curative tool against cancer. *Pharmaceutics*. 2022; 14(10): 2100. DOI: [10.3390/pharmaceutics14102100](https://doi.org/10.3390/pharmaceutics14102100)
2. Minchinton AI, and Tannock IF. Drug penetration in solid tumours. *Nat Rev Ca*. 2006; 6(8): 583-592. DOI: [10.1038/nrc1893](https://doi.org/10.1038/nrc1893)
3. St Jean AT, Zhang M, and Forbes NS. Bacterial therapies: Completing the cancer treatment toolbox. *Curr Opin Biotechnol*. 2008; 19(5): 511-517. DOI: [10.1016/j.copbio.2008.08.004](https://doi.org/10.1016/j.copbio.2008.08.004)
4. Keung EZ, Fairweather M, and Raut CP. Surgical management of metastatic disease. *Surg Clin North Am*. 2016; 96(5): 1175-1192. DOI: [10.1016/j.suc.2016.05.010](https://doi.org/10.1016/j.suc.2016.05.010)
5. Dutt S, Ahmed MM, Loo BW Jr, and Strober S. Novel radiation therapy paradigms and immunomodulation: Heresies and hope. *Semin Radiat Oncol*. 2020; 30(2): 194-200. DOI: [10.1016/j.semradonc.2019.12.006](https://doi.org/10.1016/j.semradonc.2019.12.006)
6. Kocakavuk E, Anderson KJ, Varn FS, Johnson KC, Amin SB, Sulman EP, et al. Radiotherapy is associated with a deletion signature that contributes to poor outcomes in patients with cancer. *Nat Genet*. 2021; 53(7): 1088-1096. DOI: [10.1038/s41588-021-00874-3](https://doi.org/10.1038/s41588-021-00874-3)
7. Saeed M, Sadr S, Gharib A, Lotfalizadeh N, Hajjafari A, Simab PA, et al. Phytosomes: A promising nanocarrier for enhanced delivery of herbal compounds in cancer therapy. *J Lab Anim Res*. 2022; 1(1): 26-32. DOI: [10.58803/jlar.v1i1.8](https://doi.org/10.58803/jlar.v1i1.8)
8. Jing X, Yang F, Shao C, Wei K, Xie M, Shen H, et al. Role of hypoxia in cancer therapy by regulating the tumor microenvironment. *Mol Can*. 2019; 18(1): 157. DOI: [10.1186/s12943-019-1089-9](https://doi.org/10.1186/s12943-019-1089-9)
9. Rohwer N, and Cramer T. Hypoxia-mediated drug resistance: Novel insights on the functional interaction of HIFs and cell death pathways. *Drug Resist Updat*. 2011; 14(3): 191-201. DOI: [10.1016/j.drug.2011.03.001](https://doi.org/10.1016/j.drug.2011.03.001)
10. Asouli A, Sadr S, Mohebalian H, and Borji H. Anti-tumor effect of protoscolex hydatid cyst somatic antigen on inhibition cell growth of K562. *Acta Parasitol*. 2023; 68(2): 385-392. DOI: [10.1007/s11686-023-00680-3](https://doi.org/10.1007/s11686-023-00680-3)
11. Sadr S, Yousefzani Z, Simab PA, Jafari Rahbar Alizadeh A, Lotfalizadeh N, et al. *Trichinella spiralis* as a potential antitumor agent: An update. *World Vet J*. 2023; 13(1): 65-74. DOI: [10.54203/scil.2023.wvj7](https://doi.org/10.54203/scil.2023.wvj7)
12. Sadr S, Ghiassi S, Lotfalizadeh N, Simab PA, Hajjafari A, and Borji H. Antitumor mechanisms of molecules secreted by *Trypanosoma cruzi* in colon and breast cancer: A review. *Anti-cancer Agt Med Chem*. 2023; 23(15): 1710-1721 DOI: [10.2174/1871520623666230529141544](https://doi.org/10.2174/1871520623666230529141544)
13. Asouli A, Sadr S, Mohebalian H, and Borji H. Anti-tumor effect of protoscolex hydatid cyst somatic antigen on inhibition cell growth of K562. *Acta Parasitol*. 2023; 68: 385-392. DOI: [10.1007/s11686-023-00680-3](https://doi.org/10.1007/s11686-023-00680-3)
14. Sadr S, and Borji H. *Echinococcus granulosus* as a promising therapeutic agent against triple-negative breast cancer. *Curr Cancer Ther Rev*. 2023; 19(4): 292-297. DOI: [10.2174/1573394719666230427094247](https://doi.org/10.2174/1573394719666230427094247)
15. Dang LH, Bettgowda C, Huso DL, Kinzler KW, and Vogelstein B. Combination bacteriolytic therapy for the treatment of experimental tumors. *Proc Natl Acad Sci U S A*. 2001; 98(26): 15155-15160. DOI: [10.1073/pnas.251543698](https://doi.org/10.1073/pnas.251543698)
16. Badasyan I, and Nushikyan RV. Investigation of salmonellosis during and after the COVID-19 pandemic (2020-2023). *Res Biotechnol Environ Sci*. 2023; 2(2): 30-34. DOI: [10.58803/rbes.v2i2.12](https://doi.org/10.58803/rbes.v2i2.12)
17. Akoachere JF, Tanih NF, Ndip LM, and Ndip RN. Phenotypic characterization of *Salmonella typhimurium* isolates from food-animals and abattoir drains in Buea, Cameroon. *J H Popul Nutr*. 2009; 27(5): 612-618. DOI: [10.3329/jhpn.v27i5.3637](https://doi.org/10.3329/jhpn.v27i5.3637)
18. Liang K, Liu Q, Li P, Luo H, Wang H, and Kong Q. Genetically engineered *Salmonella Typhimurium*: Recent advances in cancer therapy. *Can Lett*. 2019; 448: 168-181. DOI: [10.1016/j.canlet.2019.01.037](https://doi.org/10.1016/j.canlet.2019.01.037)
19. Chen F, Zang Z, Chen Z, Cui L, Chang Z, Ma A, et al. Nanophotosensitizer-engineered *Salmonella* bacteria with hypoxia targeting and photothermal-assisted mutual bioaccumulation for solid tumor therapy. *Biomaterials*. 2019; 214: 119226. DOI: [10.1016/j.biomaterials.2019.119226](https://doi.org/10.1016/j.biomaterials.2019.119226)
20. Wei MQ, Ellem KA, Dunn P, West MJ, Bai CX, and Vogelstein B. Facultative or obligate anaerobic bacteria have the potential for multimodality therapy of solid tumours. *Eur J Cancer*. 2007; 43(3): 490-496. DOI: [10.1016/j.ejca.2006.10.005](https://doi.org/10.1016/j.ejca.2006.10.005)
21. Jalal K, Khan K, Hassam M, Abbas MN, Uddin R, Khusro A, et al. Identification of a novel therapeutic target against XDR *Salmonella typhi* H58 using genomics driven approach followed up by natural products virtual screening. *Microorganisms*. 2021; 9(12): 2512. DOI: [10.3390/microorganisms9122512](https://doi.org/10.3390/microorganisms9122512)
22. Mi Z, Guo L, Liu P, Qi Y, Feng Z, Liu J, et al. Trojan horse *Salmonella* enabling tumor homing of silver nanoparticles via neutrophil infiltration for synergistic tumor therapy and enhanced biosafety. *Nano Lett*. 2020; 21(1): 414-423. DOI: [10.1021/acs.nanolett.0c03811](https://doi.org/10.1021/acs.nanolett.0c03811)
23. Zhang Q, Feng Y, and Kennedy D. Multidrug-resistant cancer cells and cancer stem cells hijack cellular systems to circumvent systemic therapies, can natural products reverse this?. *Cell Mol Life Sci*. 2017; 74: 777-801. DOI: [10.1007/s00018-016-2362-3](https://doi.org/10.1007/s00018-016-2362-3)
24. Zheng JH, and Min JJ. Targeted cancer therapy using engineered *Salmonella typhimurium*. *Chonnam Med J*. 2016; 52(3): 173-184. DOI: [10.4068/cmj.2016.52.3.173](https://doi.org/10.4068/cmj.2016.52.3.173)
25. Yong L, Tang S, Yu H, Zhang H, Zhang Y, Wan Y, et al. The role of hypoxia-inducible factor-1 alpha in multidrug-resistant breast cancer. *Front Oncol*. 2022; 12: 964934. DOI: [10.3389/fonc.2022.964934](https://doi.org/10.3389/fonc.2022.964934)
26. Aganja RP, Sivasankar C, Senevirathne A, and Lee JH. Salmonella as a Promising curative tool against cancer. *Pharmaceutics*. 2022; 14(10): 2100. DOI: [10.3390/pharmaceutics14102100](https://doi.org/10.3390/pharmaceutics14102100)
27. Semenov AV, van Overbeek L, Termorshuizen AJ, and van Bruggen AH. Influence of aerobic and anaerobic conditions on survival of *Escherichia coli* O157:H7 and *Salmonella enterica* serovar *Typhimurium* in Luria-Bertani broth, farm-yard manure and slurry. *J Environ Manage*. 2011; 92(3): 780-787. DOI: [10.1016/j.jenvman.2010.10.031](https://doi.org/10.1016/j.jenvman.2010.10.031)
28. Nguyen VH, and Min J-J. *Salmonella*-mediated cancer therapy: Roles and potential. *Nucl Med Mol Imaging*. 2017; 51(2): 118-126. DOI: [10.1007/s13139-016-0415-z](https://doi.org/10.1007/s13139-016-0415-z)
29. Lowerison MR, Huang C, Lucien F, Chen S, and Song P. Ultrasound localization microscopy of renal tumor xenografts in chicken embryo is correlated to hypoxia. *Sci Rep*. 2020; 10(1): 2478. DOI: [10.1038/s41598-020-59338-z](https://doi.org/10.1038/s41598-020-59338-z)
30. Hayek I, Schatz V, Bogdan C, Jantsch J, and Lührmann A. Mechanisms controlling bacterial infection in myeloid cells under hypoxic conditions. *Cell Mol Life Sci*. 2021; 78: 1887-1907. DOI: [10.1007/s00018-020-03684-8](https://doi.org/10.1007/s00018-020-03684-8)

31. Viillard C, and Larrivee B. Tumor angiogenesis and vascular normalization: Alternative therapeutic targets. *Angiogenesis*. 2017; 20(4): 409-426. DOI: [10.1007/s10456-017-9562-9](https://doi.org/10.1007/s10456-017-9562-9)
32. Murakami T, Hiroshima Y, Matsuyama R, Homma Y, Hoffman RM, and Endo I. Role of the tumor microenvironment in pancreatic cancer. *Ann gastroenterol surg*. 2019; 3(2): 130-137. DOI: [10.1002/ags3.12225](https://doi.org/10.1002/ags3.12225)
33. Camacho EM, Mesa-Pereira B, Medina C, Flores A, and Santero E. Engineering *Salmonella* as intracellular factory for effective killing of tumour cells. *Sci Rep*. 2016; 6(1): 30591. DOI: [10.1038/srep30591](https://doi.org/10.1038/srep30591)
34. Liu F, Zhang L, Hoffman RM, and Zhao M. Vessel destruction by tumor-targeting *Salmonella typhimurium* A1-R is enhanced by high tumor vascularity. *Cell Cycle*. 2010; 9(22): 4518-4524. DOI: [10.4161/cc.9.22.13744](https://doi.org/10.4161/cc.9.22.13744)
35. Das R, and Fernandez JG. Biomaterials for mimicking and modelling tumor microenvironment. *Microfluidics and biosensors in cancer research: Applications in cancer modeling and theranostics*. *Adv Exp Med Biol*. 2022; 1379: 139-170. DOI: [10.1007/978-3-031-04039-9_6](https://doi.org/10.1007/978-3-031-04039-9_6)
36. Sullivan MR, Danai LV, Lewis CA, Chan SH, Gui DY, Kunchok T, et al. Quantification of microenvironmental metabolites in murine cancers reveals determinants of tumor nutrient availability. *Elife*. 2019; 8: e44235. DOI: [10.7554/eLife.44235](https://doi.org/10.7554/eLife.44235)
37. Pavlova NN, Zhu J, and Thompson CB. The hallmarks of cancer metabolism: Still emerging. *Cell Metab*. 2022; 34(3): 355-377. DOI: [10.1016/j.cmet.2022.01.007](https://doi.org/10.1016/j.cmet.2022.01.007)
38. Felgner S, Kocijancic D, Frahm M, and Weiss S. Bacteria in cancer therapy: Renaissance of an old concept. *Int J Microbiol*. 2016; 2016: 8451728. DOI: [10.1155/2016/8451728](https://doi.org/10.1155/2016/8451728)
39. Jawalagatti V, Kirthika P, and Lee JH. Targeting primary and metastatic tumor growth in an aggressive breast cancer by engineered tryptophan auxotrophic *Salmonella typhimurium*. *Mol Ther Oncolytics*. 2022; 25: 350-363. DOI: [10.1016/j.omto.2022.05.004](https://doi.org/10.1016/j.omto.2022.05.004)
40. Nguyen VH, and Min JJ. *Salmonella*-Mediated Cancer Therapy: Roles and Potential. *Nucl Med Mol Imaging*. 2017; 51(2): 118-126. DOI: [10.1007/s13139-016-0415-z](https://doi.org/10.1007/s13139-016-0415-z)
41. Ganai S, Arenas RB, Sauer JP, Bentley B, and Forbes NS. In tumors *Salmonella* migrate away from vasculature toward the transition zone and induce apoptosis. *Cancer Gene Ther*. 2011; 18(7): 457-466. DOI: [10.1038/cgt.2011.10](https://doi.org/10.1038/cgt.2011.10)
42. Toley BJ, and Forbes NS. Motility is critical for effective distribution and accumulation of bacteria in tumor tissue. *Integr Biol*. 2012; 4(2): 165-176. DOI: [10.1039/c2ib00091a](https://doi.org/10.1039/c2ib00091a)
43. Stritzker J, Weibel S, Seubert C, Gotz A, Tresch A, van Rooijen N, et al. Enterobacterial tumor colonization in mice depends on bacterial metabolism and macrophages but is independent of chemotaxis and motility. *Int J Med Microbiol*. 2010; 300(7): 449-456. DOI: [10.1016/j.ijmm.2010.02.004](https://doi.org/10.1016/j.ijmm.2010.02.004)
44. Zhang X, Yu D, Wu D, Gao X, Shao F, Zhao M, et al. Tissue-resident Lachnospiraceae family bacteria protect against colorectal carcinogenesis by promoting tumor immune surveillance. *Cell Host Microbe*. 2023; 31(3): 418-432 e8. DOI: [10.1016/j.chom.2023.01.013](https://doi.org/10.1016/j.chom.2023.01.013)
45. Mi Z, Feng ZC, Li C, Yang X, Ma MT, and Rong PF. *Salmonella*-mediated cancer therapy: An Innovative therapeutic strategy. *J Cancer*. 2019; 10(20): 4765-4776. DOI: [10.7150/jca.32650](https://doi.org/10.7150/jca.32650)
46. Badie F, Ghandali M, Tabatabaei SA, Safari M, Khorshidi A, Shayestehpour M, et al. Use of *Salmonella* bacteria in cancer therapy: Direct, drug delivery and combination approaches. *Front Oncol*. 2021; 11: 624759. DOI: [10.3389/fonc.2021.624759](https://doi.org/10.3389/fonc.2021.624759)
47. Hoffman RM, and Zhao M. Methods for the development of tumor-targeting bacteria. *Expert Opin Drug Discov*. 2014; 9(7): 741-750. DOI: [10.1517/17460441.2014.916270](https://doi.org/10.1517/17460441.2014.916270)
48. Chang WW, and Lee CH. *Salmonella* as an innovative therapeutic antitumor agent. *Int J Mol Sci*. 2014; 15(8): 14546-14554. DOI: [10.3390/ijms150814546](https://doi.org/10.3390/ijms150814546)
49. Lee C, Lin S, Liu J, Chang W, Hsieh J, and Wang W. *Salmonella* induce autophagy in melanoma by the downregulation of AKT/mTOR pathway. *Gene Ther*. 2014; 21(3): 309-316. DOI: [10.1038/gt.2013.86](https://doi.org/10.1038/gt.2013.86)
50. Lee CH, Lin ST, Liu JJ, Chang WW, Hsieh JL, and Wang WK. *Salmonella* induce autophagy in melanoma by the downregulation of AKT/mTOR pathway. *Gene Ther*. 2014; 21(3): 309-316. DOI: [10.1038/gt.2013.86](https://doi.org/10.1038/gt.2013.86)
51. Tsao YT, Kuo CY, Cheng SP, and Lee CH. Downregulations of AKT/mTOR signaling pathway for *Salmonella*-mediated suppression of matrix metalloproteinases-9 expression in mouse tumor models. *Int J Mol Sci*. 2018; 19(6): 1630. DOI: [10.3390/ijms19061630](https://doi.org/10.3390/ijms19061630)
52. Jiang X, Wang J, Deng X, Xiong F, Zhang S, Gong Z, et al. The role of microenvironment in tumor angiogenesis. *J Exp Clin Cancer Res*. 2020; 39(1): 1-19. DOI: [10.1186/s13046-020-01709-5](https://doi.org/10.1186/s13046-020-01709-5)
53. Tu DG, Chang WW, Lin ST, Kuo CY, Tsao YT, and Lee CH. *Salmonella* inhibits tumor angiogenesis by downregulation of vascular endothelial growth factor. *Oncotarget*. 2016; 7(25): 37513-37523. DOI: [10.18632/oncotarget.7038](https://doi.org/10.18632/oncotarget.7038)
54. Wang WK, Chen MC, Leong HF, Kuo YL, Kuo CY, and Lee CH. Connexin 43 suppresses tumor angiogenesis by down-regulation of vascular endothelial growth factor via hypoxic-induced factor-1alpha. *Int J Mol Sci*. 2014; 16(1): 439-451. DOI: [10.3390/ijms16010439](https://doi.org/10.3390/ijms16010439)
55. Kiyuna T, Tome Y, Uehara F, Murakami T, Zhang Y, Zhao M, et al. Tumor-targeting *Salmonella typhimurium* A1-R inhibits osteosarcoma angiogenesis in the *in vivo* gelfoam(R) assay visualized by color-coded imaging. *Anti-cancer Res*. 2018; 38(1): 159-164. DOI: [10.21873/anticancerres.12203](https://doi.org/10.21873/anticancerres.12203)
56. Li S, Yue H, Wang S, Li X, Wang X, Guo P, et al. Advances of bacteria-based delivery systems for modulating tumor microenvironment. *Adv Drug Deliv Rev*. 2022; 188: 114444. DOI: [10.1016/j.addr.2022.114444](https://doi.org/10.1016/j.addr.2022.114444)
57. Kaimala S, Al-Sbiei A, Cabral-Marques O, Fernandez-Cabezudo MJ, and Al-Ramadi BK. Attenuated bacteria as immunotherapeutic tools for cancer treatment. *Front Oncol*. 2018; 8: 136. DOI: [10.3389/fonc.2018.00136](https://doi.org/10.3389/fonc.2018.00136)
58. Lee CH, Hsieh JL, Wu CL, Hsu HC, and Shiau AL. B cells are required for tumor-targeting *Salmonella* in host. *Appl Microbiol Biotechnol*. 2011; 92(6): 1251-1260. DOI: [10.1007/s00253-011-3386-0](https://doi.org/10.1007/s00253-011-3386-0)
59. Grille S, Moreno M, Bascuas T, Marques JM, Munoz N, Lens D, et al. *Salmonella enterica* serovar *Typhimurium* immunotherapy for B-cell lymphoma induces broad anti-tumour immunity with therapeutic effect. *Immunology*. 2014; 143(3): 428-437. DOI: [10.1111/imm.12320](https://doi.org/10.1111/imm.12320)
60. Galdiero MR, Bonavita E, Barajon I, Garlanda C, Mantovani A, and Jaillon S. Tumor associated macrophages and neutrophils in cancer. *Immunobiol*. 2013; 218(11): 1402-1410. DOI: [10.1016/j.imbio.2013.06.003](https://doi.org/10.1016/j.imbio.2013.06.003)
61. Duan B, Shao L, Liu R, Msuthwana P, Hu J, and Wang C. *Lactobacillus rhamnosus* GG defense against *Salmonella enterica* serovar *Typhimurium* infection through modulation of M1 macrophage polarization. *Microb Pathog*. 2021; 156: 104939. DOI: [10.1016/j.micpath.2021.104939](https://doi.org/10.1016/j.micpath.2021.104939)
62. Mantovani A, Sica A, Sozzani S, Allavena P, Vecchi A, and Locati M. The chemokine system in diverse forms of macrophage activation and polarization. *Trends Immunol*. 2004; 25(12): 677-686. DOI: [10.1016/j.it.2004.09.015](https://doi.org/10.1016/j.it.2004.09.015)
63. Kaimala S, Mohamed YA, Nader N, Issac J, Elkord E, Chouaib S, et al. *Salmonella*-mediated tumor regression involves targeting of tumor myeloid suppressor cells causing a shift to M1-like phenotype and reduction in suppressive capacity. *Cancer Immunology, IO*. 2014; 63: 587-399. DOI: [10.1007/s00262-014-1543-x](https://doi.org/10.1007/s00262-014-1543-x)
64. Josefowicz SZ, Lu LF, and Rudensky AY. Regulatory T cells: Mechanisms of differentiation and function. *Annu Rev Immunol*. 2012; 30: 531-564. DOI: [10.1146/annurev.immunol.25.022106.141623](https://doi.org/10.1146/annurev.immunol.25.022106.141623)
65. Hong EH, Chang SY, Lee BR, Pyun AR, Kim JW, Kweon MN, et al. Intratumoral injection of attenuated *Salmonella* vaccine can induce tumor microenvironmental shift from immune suppressive to immunogenic. *Vaccine*. 2013; 31(10): 1377-1384. DOI: [10.1016/j.vaccine.2013.01.006](https://doi.org/10.1016/j.vaccine.2013.01.006)
66. Liu T, and Chopra AK. An enteric pathogen *Salmonella enterica* serovar *Typhimurium* suppresses tumor growth by downregulating CD44high and CD4T regulatory (Treg) cell expression in mice: The critical role of lipopolysaccharide and Braun lipoprotein in modulating tumor growth. *Cancer Gene Ther*. 2010; 17(2): 97-108. DOI: [10.1038/cgt.2009.58](https://doi.org/10.1038/cgt.2009.58)
67. Varela-Vazquez A, Guitian-Caamano A, Carpintero-Fernandez P, Fonseca E, Sayedyahosseini S, Aasen T, et al. Emerging functions and clinical prospects of connexins and pannexins in melanoma. *Biochim Biophys Acta Rev Cancer*. 2020; 1874(1): 188380. DOI:

- 10.1016/j.bbcan.2020.188380
68. Saccheri F, Pozzi C, Avogadri F, Barozzi S, Faretta M, Fusi P, et al. Bacteria-induced gap junctions in tumors favor antigen cross-presentation and antitumor immunity. *Sci Transl Med.* 2010; 2(44): 44-57. DOI: [10.1126/scitranslmed.3000739](https://doi.org/10.1126/scitranslmed.3000739)
 69. DeNardo DG, and Ruffell B. Macrophages as regulators of tumour immunity and immunotherapy. *Nat Rev Immunol.* 2019; 19(6): 369-382. DOI: [10.1038/s41577-019-0127-6](https://doi.org/10.1038/s41577-019-0127-6)
 70. Biswas SK, Gangi L, Paul S, Schioppa T, Saccani A, Sironi M, et al. A distinct and unique transcriptional program expressed by tumor-associated macrophages (defective NF-kappaB and enhanced IRF-3/STAT1 activation). *Blood.* 2006; 107(5): 2112-2122. DOI: [10.1182/blood-2005-01-0428](https://doi.org/10.1182/blood-2005-01-0428)
 71. Boutilier AJ, and ElSawa SF. Macrophage polarization states in the tumor microenvironment. *Int J Mol Sci.* 2021; 22(13): 6995. DOI: [10.3390/ijms22136995](https://doi.org/10.3390/ijms22136995)
 72. Jorgovanovic D, Song M, Wang L, and Zhang Y. Roles of IFN- γ in tumor progression and regression: A review. *Biomark Res.* 2020; 8: 49. DOI: [10.1186/s40364-020-00228-x](https://doi.org/10.1186/s40364-020-00228-x)
 73. Chen G, Xiong W, Gu Z, Gao Y, Hou J, Long L, et al. Mannosylated engineered trichosanthin-legumain protein vaccine hydrogel for breast cancer immunotherapy. *Int J Biol Macromol.* 2022; 223: 1485-1494. DOI: [10.1016/j.ijbiomac.2022.11.045](https://doi.org/10.1016/j.ijbiomac.2022.11.045)
 74. Luo Y, Zhou H, Krueger J, Kaplan C, Lee SH, Dolman C, et al. Targeting tumor-associated macrophages as a novel strategy against breast cancer. *J Clin Invest.* 2006; 116(8): 2132-2141. DOI: [10.1172/JCI27648](https://doi.org/10.1172/JCI27648)
 75. Kong Q, and Zhang Z. Cancer-associated pyroptosis: A new license to kill tumor. *Front Immunol.* 2023; 14: 1082165. DOI: [10.3389/fimmu.2023.1082165](https://doi.org/10.3389/fimmu.2023.1082165)
 76. Deng W, Marshall NC, Rowland JL, McCoy JM, Worrall LJ, Santos AS, et al. Assembly, structure, function and regulation of type III secretion systems. *Nat Rev Microbiol.* 2017; 15(6): 323-337. DOI: [10.1038/nrmicro.2017.20](https://doi.org/10.1038/nrmicro.2017.20)
 77. Sorenson BS, Banton KL, Frykman NL, Leonard AS, and Saltzman DA. Attenuated *Salmonella typhimurium* with IL-2 gene reduces pulmonary metastases in murine osteosarcoma. *Clin Orthop Relat Res.* 2008; 466(6): 1285-1291. DOI: [10.1007/s11999-008-0243-2](https://doi.org/10.1007/s11999-008-0243-2)
 78. Loeffler M, Le'Negrate G, Krajewska M, and Reed JC. IL-18-producing *Salmonella* inhibit tumor growth. *Cancer Gene Ther.* 2008; 15(12): 787-794. DOI: [10.1038/cgt.2008.48](https://doi.org/10.1038/cgt.2008.48)
 79. Ganai S, Arenas RB, and Forbes NS. Tumour-targeted delivery of TRAIL using *Salmonella typhimurium* enhances breast cancer survival in mice. *Br J Cancer.* 2009; 101(10): 1683-1691. DOI: [10.1038/sj.bjc.6605403](https://doi.org/10.1038/sj.bjc.6605403)
 80. Loeffler M, Le'Negrate G, Krajewska M, and Reed JC. Inhibition of tumor growth using salmonella expressing Fas ligand. *J Natl Cancer Inst.* 2008; 100(15): 1113-1116. DOI: [10.1093/jnci/djn205](https://doi.org/10.1093/jnci/djn205)
 81. Loeffler M, Le'Negrate G, Krajewska M, and Reed JC. Inhibition of tumor growth using salmonella expressing Fas ligand. *J Natl Cancer Inst.* 2008; 100(15): 1113-1116. DOI: [10.1093/jnci/djn205](https://doi.org/10.1093/jnci/djn205)
 82. Chen W, Zhu Y, Zhang Z, and Sun X. Advances in *Salmonella Typhimurium*-based drug delivery system for cancer therapy. *Adv Drug Deliv.* 2022; 185: 114295. DOI: [10.1016/j.addr.2022.114295](https://doi.org/10.1016/j.addr.2022.114295)
 83. Nallar SC, Xu DQ, and Kalvakolanu DV. Bacteria and genetically modified bacteria as cancer therapeutics: Current advances and challenges. *Cytokine.* 2017; 89: 160-172. DOI: [10.1016/j.cyto.2016.01.002](https://doi.org/10.1016/j.cyto.2016.01.002)
 84. Yang M, Yang F, Chen W, Liu S, Qiu L, and Chen J. Bacteria-mediated cancer therapies: Opportunities and challenges. *Biomater Sci.* 2021; 9(17): 5732-5744. DOI: [10.1039/D1BM00634G](https://doi.org/10.1039/D1BM00634G)
 85. Ganai S, Arenas R, and Forbes N. Tumour-targeted delivery of TRAIL using *Salmonella typhimurium* enhances breast cancer survival in mice. *Br J Cancer.* 2009; 101(10): 1683-1691. DOI: [10.1038/sj.bjc.6605403](https://doi.org/10.1038/sj.bjc.6605403)
 86. Broadway KM, and Scharf BE. *Salmonella typhimurium* as an anticancer therapy: Recent advances and perspectives. *Curr Clin Microbiol.* 2019; 6: 225-239. DOI: [10.1007/s40588-019-00132-5](https://doi.org/10.1007/s40588-019-00132-5)
 87. Chen J, Yang B, Cheng X, Qiao Y, Tang B, Chen G, et al. *Salmonella*-mediated tumor-targeting TRAIL gene therapy significantly suppresses melanoma growth in mouse model. *Cancer Sci.* 2012; 103(2): 325-333. DOI: [10.1111/j.1349-7006.2011.02147.x](https://doi.org/10.1111/j.1349-7006.2011.02147.x)
 88. Kim SI, Kim S, Kim E, Hwang SY, and Yoon H. Secretion of *Salmonella* pathogenicity island 1-encoded type III secretion system effectors by outer membrane vesicles in *Salmonella enterica* serovar typhimurium. *Front Microbiol.* 2018; 9: 2810. DOI: [10.3389/fmicb.2018.02810](https://doi.org/10.3389/fmicb.2018.02810)
 89. Galan JE, and Wolf-Watz H. Protein delivery into eukaryotic cells by type III secretion machines. *Nature.* 2006; 444(7119): 567-573. DOI: [10.1038/nature05272](https://doi.org/10.1038/nature05272)
 90. Lou L, Zhang P, Piao R, and Wang Y. *Salmonella* pathogenicity island 1 (SPI-1) and its complex regulatory network. *Front Cell Infect Microbiol.* 2019; 9: 270. DOI: [10.3389/fcimb.2019.00270](https://doi.org/10.3389/fcimb.2019.00270)
 91. Liss V, Swart AL, Kehl A, Hermanns N, Zhang Y, Chikkaballi D, et al. *Salmonella enterica* remodels the host cell endosomal system for efficient intravacuolar nutrition. *Cell Host Microbe.* 2017; 21(3): 390-402. DOI: [10.1016/j.chom.2017.02.005](https://doi.org/10.1016/j.chom.2017.02.005)
 92. Rajashekar R, Liebl D, Seitz A, and Hensel M. Dynamic remodeling of the endosomal system during formation of *Salmonella*-induced filaments by intracellular *Salmonella enterica*. *Traffic.* 2008; 9(12): 2100-2116. DOI: [10.1111/j.1600-0854.2008.00821.x](https://doi.org/10.1111/j.1600-0854.2008.00821.x)
 93. Zhang K, Riba A, Nietschke M, Torow N, Repnik U, Pütz A, et al. Minimal SPI1-T3SS effector requirement for *Salmonella* enterocyte invasion and intracellular proliferation *in vivo*. *PLoS Patho.* 2018; 14(3): e1006925. DOI: [10.1371/journal.ppat.1006925](https://doi.org/10.1371/journal.ppat.1006925)
 94. Lin HH, Chen HL, Weng CC, Janapatla RP, Chen CL, and Chiu CH. Activation of apoptosis by *Salmonella* pathogenicity island-1 effectors through both intrinsic and extrinsic pathways in *Salmonella*-infected macrophages. *Microbiol Immunol Infect.* 2021; 54(4): 616-626. DOI: [10.1016/j.jmii.2020.02.008](https://doi.org/10.1016/j.jmii.2020.02.008)
 95. Al-Saafeen BH, Fernandez-Cabezudo MJ, and Al-Ramadi BK. Integration of *Salmonella* into combination cancer therapy. *Cancers.* 2021; 13(13): 3228. DOI: [10.3390/cancers13133228](https://doi.org/10.3390/cancers13133228)
 96. Mercado-Lubo R, Zhang Y, Zhao L, Rossi K, Wu X, Zou Y, et al. A *Salmonella* nanoparticle mimic overcomes multidrug resistance in tumours. *Nat Commun.* 2016; 7: 12225. DOI: [10.1038/ncomms12225](https://doi.org/10.1038/ncomms12225)
 97. Siccardi D, Mumy KL, Wall DM, Bien JD, and McCormick BA. *Salmonella enterica* serovar *Typhimurium* modulates P-glycoprotein in the intestinal epithelium. *Am J Physiol Gastrointest Liver Physiol.* 2008; 294(6): G1392-1400. DOI: [10.1152/ajpgi.00599.2007](https://doi.org/10.1152/ajpgi.00599.2007)
 98. Sundaram B, and Kanneganti TD. Advances in understanding activation and function of the NLRC4 inflammasome. *Int J Mol Sci.* 2021; 22(3): 1048. DOI: [10.3390/ijms22031048](https://doi.org/10.3390/ijms22031048)
 99. Yang J, Liu Z, and Xiao TS. Post-translational regulation of inflammasomes. *Cell mol immun.* 2017; 14(1): 65-79. DOI: [10.1038/cmi.2016.29](https://doi.org/10.1038/cmi.2016.29)
 100. Sundaram B, and Kanneganti TD. Advances in understanding activation and function of the NLRC4 inflammasome. *Int J Mol Sci.* 2021; 22(3): 1048. DOI: [10.3390/ijms22031048](https://doi.org/10.3390/ijms22031048)
 101. Christgen S, Zheng M, Kesavardhana S, Karki R, Malireddi RKS, Banoth B, et al. Identification of the PANoptosome: A molecular platform triggering pyroptosis, apoptosis, and necroptosis (PANoptosis). *Front Cell Infect Microbiol.* 2020; 10: 237. DOI: [10.3389/fcimb.2020.00237](https://doi.org/10.3389/fcimb.2020.00237)
 102. Liu X, Zhang Z, Ruan J, Pan Y, Magupalli VG, Wu H, et al. Inflammasome-activated gasdermin D causes pyroptosis by forming membrane pores. *Nature.* 2016; 535(7610): 153-158. DOI: [10.1038/nature18629](https://doi.org/10.1038/nature18629)
 103. Raymond B, Young JC, Pallett M, Endres RG, Clements A, and Frankel G. Subversion of trafficking, apoptosis, and innate immunity by type III secretion system effectors. *Trends Microbiol.* 2013; 21(8): 430-441. DOI: [10.1016/j.tim.2013.06.008](https://doi.org/10.1016/j.tim.2013.06.008)
 104. Nishikawa H, Sato E, Briones G, Chen LM, Matsuo M, Nagata Y, et al. *In vivo* antigen delivery by a *Salmonella typhimurium* type III secretion system for therapeutic cancer vaccines. *J Clin Invest.* 2006; 116(7): 1946-1954. DOI: [10.1172/JCI28045](https://doi.org/10.1172/JCI28045)
 105. Roeder E, Jellbauer S, Kohn B, Berchtold C, Partilla M, Busch DH, et al. Invasion and destruction of a murine fibrosarcoma by *Salmonella*-induced effector CD8 T cells as a therapeutic intervention against cancer. *Cancer Immunol Immunother.* 2011; 60(3): 371-380. DOI: [10.1007/s00262-010-0950-x](https://doi.org/10.1007/s00262-010-0950-x)
 106. Xiong G, Husseiny MI, Song L, Erdreich-Epstein A, Shackleford GM, Seeger RC, et al. Novel cancer vaccine based on genes of *Salmonella* pathogenicity island 2. *Int J Cancer.* 2010; 126(11): 2622-2634. DOI: [10.1002/ijc.24957](https://doi.org/10.1002/ijc.24957)
 107. Panthel K, Meinel KM, Sevil Domenech VE, Trulzsch K, and Russmann

- H. Salmonella type III-mediated heterologous antigen delivery: A versatile oral vaccination strategy to induce cellular immunity against infectious agents and tumors. *Int J Med Microbiol.* 2008; 298(1-2): 99-103. DOI: [10.1016/j.ijmm.2007.07.002](https://doi.org/10.1016/j.ijmm.2007.07.002)
108. Liang K, Liu Q, Li P, Luo H, Wang H, and Kong Q. Genetically engineered *Salmonella Typhimurium*: Recent advances in cancer therapy. *Cancer Lett.* 2019; 448: 168-181. DOI: [10.1016/j.canlet.2019.01.037](https://doi.org/10.1016/j.canlet.2019.01.037)
109. Kung YJ, Lam B, Tseng SH, MacDonald A, Tu HF, Wang S, et al. Localization of Salmonella and albumin-IL-2 to the tumor microenvironment augments anti-cancer T cell immunity. *J Biomed Sci.* 2022; 29(1): 57. DOI: [10.1186/s12929-022-00841-y](https://doi.org/10.1186/s12929-022-00841-y)
110. Sun X, Ni N, Ma Y, Wang Y, and Leong DT. Retooling cancer nanotherapeutics' entry into tumors to alleviate tumoral hypoxia. *Small.* 2020; 16(41): 2003000. DOI: [10.1002/smll.202003000](https://doi.org/10.1002/smll.202003000)
111. Frutos-Grilo E, Marsal M, Irazoki O, Barbé J, and Campoy S. The interaction of RecA with both CheA and CheW is required for Chemotaxis. *Front Microbiol.* 2020; 7(11): 583. DOI: [10.3389/fmicb.2020.00583](https://doi.org/10.3389/fmicb.2020.00583)
112. Wang L, Li Y, Liu Y, Zuo L, Li Y, Wu S, et al. Salmonella spv locus affects type I interferon response and the chemotaxis of neutrophils via suppressing autophagy. *Fish Shellfish Immunol.* 2019; 87: 721-729. DOI: [10.1016/j.fsi.2019.02.009](https://doi.org/10.1016/j.fsi.2019.02.009)
113. Singleton DC, Macann A, and Wilson WR. Therapeutic targeting of the hypoxic tumour microenvironment. *Nature Rev Clinical Oncology.* 2021; 18(12): 751-772. DOI: [10.1038/s41571-021-00539-4](https://doi.org/10.1038/s41571-021-00539-4)
114. Ma J, Sun X, Wang Y, Chen B, Qian L, and Wang Y. Fibroblast-derived CXCL12 regulates PTEN expression and is associated with the proliferation and invasion of colon cancer cells via PI3k/Akt signaling. *Cell Commun Signal.* 2019; 17(1): 119. DOI: [10.1186/s12964-019-0432-5](https://doi.org/10.1186/s12964-019-0432-5)
115. Kalia VC, Patel SK, Cho BK, Wood TK, and Lee JK. Emerging applications of bacteria as antitumor agents. *Semin Cancer Biol.* 2022; 86(Pt 2): 1014-1025. DOI: [10.1016/j.semcancer.2021.05.012](https://doi.org/10.1016/j.semcancer.2021.05.012)
116. Jimenez-Jimenez C, Moreno VM, and Vallet-Regi M. Bacteria-assisted transport of nanomaterials to improve drug delivery in cancer therapy. *Nanomaterials.* 2022; 12(2): 288. DOI: [10.3390/nano12020288](https://doi.org/10.3390/nano12020288)
117. Guo Y, Chen Y, Liu X, Min JJ, Tan W, and Zheng JH. Targeted cancer immunotherapy with genetically engineered oncolytic *Salmonella typhimurium*. *Cancer Lett.* 2020; 469: 102-110. DOI: [10.1016/j.canlet.2019.10.033](https://doi.org/10.1016/j.canlet.2019.10.033)
118. Dos Santos AMP, Ferrari RG, and Conte-Junior CA. Virulence factors in *Salmonella typhimurium*: The sagacity of a bacterium. *Curr Microbiol.* 2019; 76(6): 762-773. DOI: [10.1007/s00284-018-1510-4](https://doi.org/10.1007/s00284-018-1510-4)
119. Wang W, Xu H, Ye Q, Tao F, Wheeldon I, Yuan A, et al. Systemic immune responses to irradiated tumours via the transport of antigens to the tumour periphery by injected flagellate bacteria. *Nat Biomed Eng.* 2022; 6(1): 44-53. DOI: [10.1038/s41551-021-00834-6](https://doi.org/10.1038/s41551-021-00834-6)
120. Kiani AA, Elyasi H, Ghoreyshi S, Nouri N, Safarzadeh A, and Nafari A. Study on hypoxia-inducible factor and its roles in immune system. *Immunol Med.* 2021; 44(4): 223-236. DOI: [10.1080/25785826.2021.1910187](https://doi.org/10.1080/25785826.2021.1910187)
121. Chiu HM, Chiou WY, Hsu WJ, Wu LH, Yang MH, Tyan YC, et al. Salmonella alters heparanase expression and reduces tumor metastasis. *Int J Med Sci.* 2021; 18(13): 2981. DOI: [10.7150/ijms.60281](https://doi.org/10.7150/ijms.60281)
122. Leschner S, and Weiss S. Salmonella— allies in the fight against cancer. *J Mol Med.* 2010; 88(8): 763-773. DOI: [10.1007/s00109-010-0636-z](https://doi.org/10.1007/s00109-010-0636-z)
123. Zhao M, Yang M, Ma H, Li X, Tan X, Li S, et al. Targeted therapy with a *Salmonella typhimurium* leucine-arginine auxotroph cures orthotopic human breast tumors in nude mice. *Cancer Res.* 2006; 66(15): 7647-7652. DOI: [10.1158/0008-5472.CAN-06-0716](https://doi.org/10.1158/0008-5472.CAN-06-0716)
124. Nguyen VH, Kim HS, Ha JM, Hong Y, Choy HE, and Min JJ. Genetically engineered *Salmonella typhimurium* as an imageable therapeutic probe for cancer. *Cancer Res.* 2010; 70(1): 18-23. DOI: [10.1158/0008-5472.CAN-09-3453](https://doi.org/10.1158/0008-5472.CAN-09-3453)
125. Zheng JH, Nguyen VH, Jiang SN, Park SH, Tan W, Hong SH, et al. Two-step enhanced cancer immunotherapy with engineered *Salmonella typhimurium* secreting heterologous flagellin. *Sci Trans Med.* 2017; 9(376): eaak9537. DOI: [10.1126/scitranslmed.aak9537](https://doi.org/10.1126/scitranslmed.aak9537)
126. Al-Ramadi BK, Fernandez-Cabezudo MJ, El-Hasasna H, Al-Salam S, Bashir G, and Chouaib S. Potent anti-tumor activity of systemically-administered IL2-expressing *Salmonella* correlates with decreased angiogenesis and enhanced tumor apoptosis. *Clin Immunol.* 2009; 130(1): 89-97. DOI: [10.1016/j.clim.2008.08.021](https://doi.org/10.1016/j.clim.2008.08.021)
127. Zhang Y, Tome Y, Suetsugu A, Zhang L, Zhang N, Hoffman RM, et al. Determination of the optimal route of administration of *Salmonella typhimurium* A1-R to target breast cancer in nude mice. *Anti-cancer Res.* 2012; 32(7): 2501-2508.
128. Li X, Li Y, Wang B, Ji K, Liang Z, Guo B, et al. Delivery of the co-expression plasmid pEndo-Si-Stat3 by attenuated *Salmonella serovar typhimurium* for prostate cancer treatment. *J Cancer Res Clin Oncol.* 2013; 139(6): 971-980. DOI: [10.1007/s00432-013-1398-0](https://doi.org/10.1007/s00432-013-1398-0)
129. Schatten H. Immunodiagnosics and immunotherapy possibilities for prostate cancer. *Adv Exp Med Biol.* 2018; 1096: 185-94. DOI: [10.1007/978-3-319-99286-0_10](https://doi.org/10.1007/978-3-319-99286-0_10)
130. Wang CZ, Kazmierczak RA, and Eisenstark A. Strains, mechanism, and perspective: Salmonella-based cancer therapy. *Int J Microbiol.* 2016; 5678702. DOI: [10.1155/2016/5678702](https://doi.org/10.1155/2016/5678702)
131. Pangilinan CR, and Lee CH. Salmonella-based targeted cancer therapy: Updates on a promising and innovative tumor immunotherapeutic strategy. *Biomed.* 2019; 7(2): 36. DOI: [10.3390/biomedicines7020036](https://doi.org/10.3390/biomedicines7020036)
132. Ebrahimzadeh S, Ahangari H, Soleimanian A, Hosseini K, Ebrahimi V, Ghasemnejad T, et al. Colorectal cancer treatment using bacteria: focus on molecular mechanisms. *BMC Microbiol.* 2021; 21(1): 218. DOI: [10.1186/s12866-021-02274-3](https://doi.org/10.1186/s12866-021-02274-3)
133. Deng J, Guo Y, Jiang Z, Yang M, Li H, and Wang J. Enhancement of ovarian cancer chemotherapy by delivery of multidrug-resistance gene small interfering RNA using tumor targeting *Salmonella*. *J Obstet Gynaecol Res.* 2015; 41(4): 615-622. DOI: [10.1111/jog.12598](https://doi.org/10.1111/jog.12598)
134. Fu W, Lan H, Li S, Han X, Gao T, and Ren D. Synergistic antitumor efficacy of suicide/ePNP gene and 6-methylpurine 2'-deoxyriboside via *Salmonella* against murine tumors. *Cancer Gene Ther.* 2008; 15(7): 474-484. DOI: [10.1038/cgt.2008.19](https://doi.org/10.1038/cgt.2008.19)
135. Phan T, Nguyen VH, D'Alincourt MS, Manuel ER, Kaltcheva T, Tsai W, et al. Salmonella-mediated therapy targeting indoleamine 2, 3-dioxygenase 1 (IDO) activates innate immunity and mitigates colorectal cancer growth. *Cancer Gene Ther.* 2020; 27(3-4): 235-245. DOI: [10.1038/s41417-019-0089-7](https://doi.org/10.1038/s41417-019-0089-7)
136. Pangilinan CR, Wu LH, and Lee CH. Salmonella impacts tumor-induced macrophage polarization, and inhibits SNAIL-mediated metastasis in melanoma. *Cancers.* 2021; 13(12): 2894. DOI: [10.3390/cancers13122894](https://doi.org/10.3390/cancers13122894)
137. Yoon W, Park YC, Kim J, Chae YS, Byeon JH, Min SH, et al. Application of genetically engineered *Salmonella typhimurium* for interferon-gamma-induced therapy against melanoma. *Eur J Cancer.* 2017; 70: 48-61. DOI: [10.1016/j.ejca.2016.10.010](https://doi.org/10.1016/j.ejca.2016.10.010)
138. Oladejo M, Paulishak W, and Wood L. Synergistic potential of immune checkpoint inhibitors and therapeutic cancer vaccines. *Semin Cancer Biol.* 2022; 88: 81-95. DOI: [10.1016/j.semcancer.2022.12.003](https://doi.org/10.1016/j.semcancer.2022.12.003)
139. Chen Y, Liu X, Guo Y, Wang J, Zhang D, Mei Y, et al. Genetically engineered oncolytic bacteria as drug delivery systems for targeted cancer theranostics. *Acta Biomaterialia.* 2021; 124: 72-87. DOI: [10.1016/j.actbio.2021.02.006](https://doi.org/10.1016/j.actbio.2021.02.006)
140. Zhang Y, Sun X, Wang Q, Xu J, Dong F, Yang S, et al. Multicopy chromosomal integration using CRISPR-associated transposases. *ACS Synth Biol.* 2020; 9(8): 1998-2008. DOI: [10.1021/acssynbio.0c00073](https://doi.org/10.1021/acssynbio.0c00073)
141. Zhou S, Gravekamp C, Bermudes D, and Liu K. Tumour-targeting bacteria engineered to fight cancer. *Nat Rev Cancer.* 2018; 18(12): 727-743. DOI: [10.1038/s41568-018-0070-z](https://doi.org/10.1038/s41568-018-0070-z)
142. Sedighi M, Zahedi Bialvaei A, Hamblin MR, Ohadi E, Asadi A, Halajzadeh M, et al. Therapeutic bacteria to combat cancer; current advances, challenges, and opportunities. *Cancer Med.* 2019; 8(6): 3167-3181. DOI: [10.1002/cam4.2148](https://doi.org/10.1002/cam4.2148)
143. Forbes NS, Coffin RS, Deng L, Evgin L, Fiering S, Giacalone M, et al. White paper on microbial anti-cancer therapy and prevention. *J Immunother Cancer.* 2018; 6(1): 78. DOI: [10.1186/s40425-018-0381-3](https://doi.org/10.1186/s40425-018-0381-3)
144. Geller LT, Barzily-Rokni M, Danino T, Jonas OH, Shental N, Nejman D,

- et al. Potential role of intratumor bacteria in mediating tumor resistance to the chemotherapeutic drug gemcitabine. *Science*. 2017; 357(6356): 1156-1160. DOI: [10.1126/science.aah5043](https://doi.org/10.1126/science.aah5043)
145. Sharma P, and Allison JP. The future of immune checkpoint therapy. *Sci*. 2015; 348(6230): 56-61. DOI: [10.1126/science.aaa8172](https://doi.org/10.1126/science.aaa8172)
146. Zou W, Wolchok JD, and Chen L. PD-L1 (B7-H1) and PD-1 pathway blockade for cancer therapy: Mechanisms, response biomarkers, and combinations. *Sci Trans Med*. 2016; 8(328): 328rv4-328rv4. DOI: [10.1126/scitranslmed.aad7118](https://doi.org/10.1126/scitranslmed.aad7118)
147. Rock CL, Thomson C, Gansler T, Gapstur SM, McCullough ML, Patel AV, et al. American cancer society guideline for diet and physical activity for cancer prevention. *CA Cancer J Clin*. 2020; 70(4): 245-271. DOI: [10.3322/caac.21591](https://doi.org/10.3322/caac.21591)